

This Page Is Inserted by IFW Operations
and is not a part of the Official Record

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

IMAGES ARE BEST AVAILABLE COPY.

**As rescanning documents *will not* correct images,
please do not report the images to the
Image Problem Mailbox.**

(19) World Intellectual Property Organization
International Bureau(43) International Publication Date
23 May 2002 (23.05.2002)

PCT

(10) International Publication Number
WO 02/40696 A2(51) International Patent Classification⁷: **C12P 21/02**

[TH/TH]; 234/171 Siriwatana Nives Mahidol Road, Nong Hoi, Muang Chiang Mai 50000 (TH).

(21) International Application Number: PCT/EP01/12920

(22) International Filing Date:
8 November 2001 (08.11.2001)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
0027782.2 14 November 2000 (14.11.2000) GB(71) Applicant (for all designated States except US):
BOEHRINGER INGELHEIM INTERNATIONAL GMBH [DE/DE]; 55216 Ingelheim / Rhein (DE).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **WERNER, Rolf-Günther** [DE/DE]; Hugo-Haering-Strasse 72, 88400 Biberach an der Riss (DE). **GOETZ, Friedrich** [DE/DE]; Beim Herbstenhof 31, 72076 Tübingen (DE). **TAYAPI-WATANA, Chatchai** [TH/TH]; 369/73 Soi Pradoo 1, Janava Rd., BKK (TH). **MANOSROI, Jiradej** [TH/TH]; 234/171 Siriwatana Nives Mahidol Road, Nong Hoi, Muang Chiang Mai 50000 (TH). **MANOSROI, Aranya**

(81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

— without international search report and to be republished upon receipt of that report

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: METHODS FOR LARGE SCALE PROTEIN PRODUCTION IN PROKARYOTES

(57) Abstract: The invention belongs to the field of protein production in prokaryotic cells. The invention relates to methods for the production of recombinant DNA-derived heterologous protein in prokaryotic cells, wherein said heterologous protein is secreted extracellularly as an active and correctly folded protein, and the prokaryotic cell contains and expresses a vector comprising the DNA coding for said heterologous protein operably linked to the DNA coding for the signal peptide OmpA.

WO 02/40696 A2

Methods for large scale protein production in prokaryotes

The invention belongs to the field of protein production in prokaryotic cells.

The invention relates to methods for the production of recombinant DNA-derived heterologous protein in prokaryotic cells, wherein said heterologous protein is secreted extracellularly as an active and correctly folded protein, and the prokaryotic cell contains and expresses a vector comprising the DNA coding for said heterologous protein operably linked to the DNA coding for the signal peptide OmpA.

Background art

Prokaryotic expression systems for heterologous proteins are commonly used for proteins which do not require mammalian glycosylation patterns as they provide a cheap way of producing large quantities of said protein. The formation of highly aggregated protein or inclusion bodies can be commonly found in high-level expression of many heterologous proteins in *E.coli*. One way of protein production is via inclusion bodies which develop in cytoplasm. Cell wall and outer membrane components of the prokaryotic cells used for production (e.g. *E.coli*) usually contaminate the cell lysate containing the heterologous protein when said inclusion bodies are prepared by low-speed centrifugation. The outer membrane component can be eliminated by selective extraction with detergents and low concentrations of either urea or guanidine-HCl.

One example of such a heterologous protein is a tPA derivative.

Tissue plasminogen activator (tPA) is a polypeptide containing 527 amino acid residues (27) with a molecular mass of 72 kDa. The molecule is divided into five structural domains. Nearby the N-terminal region is a looped finger domain, which is followed by a growth factor domain. Two similar domains, kringle 1 and kringle 2, are following. Both finger and kringle 2 domains bind specifically to the fibrin clots thereby accelerating tPA protein activation of bound plasminogen. Downstream of kringle 2 is the serine protease, with its catalytic site located at the C-terminus. The serine protease is responsible for converting plasminogen to plasmin a reaction important in the homeostasis of fibrin formation and clot dissolution. The correct folding of tPA requires the correct pairing of 17 disulfide bridges in the molecule (1).

Clinically, tPA is a thrombolytic agent of choice for the treatment of acute myocardial infarction. It has the advantage of causing no side effects on systemic haemorrhaging and fibrinogen depletion (7). Bowes melanoma cells were first used as a source in tPA production for

therapeutic purposes (12). Since a consistent process with efficient production of highly purified protein in good yield is required for clinical use, the construction of full-length recombinant-tPA (r-tPA) progressed to mammalian cells. Chinese hamster ovary cells were transfected with the tPA gene to synthesize the r-tPA (8, 22). The recombinant product produced by a mammalian fermentation system was harvested from the culture medium. Attracted by simplicity and economy of production, a number of efforts in producing r-tPA from bacteria, especially from *Escherichia coli*, were investigated (10, 13, 30). Regarding the low yield and the formation of inclusion bodies, which resulted in misfolding and in an inactive enzyme, numerous strategies have been proposed to overcome these problems. The major criterion is to synthesize the smallest molecule, which is still active instead of full-length tPA.

Several deletion-mutant variants including kringle 2 plus serine protease (K2S) were considered. However, the enzymatic activity of the recombinant-K2S (r-K2S) was obtained only when refolding processes of purified inclusion bodies from cytoplasmic compartment were achieved (16, 29). In order to avoid the cumbersome refolding processes and periplasmic protein delivery, special bacterial expression systems were exploited (6, 31). Despite periplasmic expression of tPA, overexpression led to inactive aggregates, even in the relatively high oxidizing condition in the periplasm.

In the prior art, there are a few descriptions of methods for the preparation of recombinant K2S in *E. coli*. However, there is no disclosure of a method leading to a cost effective method for large scale production of biologically active K2S.

Obukowicz et al. (25) expressed and purified r-K2S from periplasmic space. The obvious disadvantage of this method was an extra periplasmic extraction step, which is not suitable for large scale production.

Saito et al. (29) disclose the cytoplasmic expression of r-K2S. The authors used an in vivo renaturation processes for the expressed r-K2S, which was purified from the cytoplasmic space of *E. coli* as inclusion body. Boehringer Mannheim use a similar cumbersome denaturing/refolding process involving the steps of cell digestion, solubilization under denaturing and reducing conditions and reactivation under oxidizing conditions in the presence of GSH/GSSG which is not cost effective and requires mutation of the amino acid sequence (24).

In 1991, Waldenström et al. (34) constructed a vector (pEZZK2P) for the secretion of kringle 2 plus serine protease domain to *E. coli* culture supernatant. Hydroxylamine was used to remove the ZZ fusion peptide from IgG-Sepharose purified fraction. The cleavage agent hydroxylamine

required modification of the cleavage sites of kringle 2 plus serine protease (Asn¹⁷⁷ → Ser and Asn¹⁸⁴ → Gln) thus to protect it from hydroxylamine digestion. However, the resulting non-native, not properly folded K2S molecule is not suitable for therapeutic purposes. The unusual sequence may even activate the human immune system.

The problem underlying the present invention was thus to provide a commercially applicable method for large scale production of heterologous proteins, e.g. K2S, wherein the heterologous protein is secreted in its biologically active form into the culture supernatant.

Description of the invention

The problem was solved within the scope of the claims and specification of the present invention.

The use of the singular or plural in the claims or specification is in no way intended to be limiting and also includes the other form.

The invention relates to a method for the production of recombinant DNA-derived heterologous protein in prokaryotic cells, wherein said heterologous protein is secreted extracellularly as an active and correctly folded protein, characterized in that the prokaryotic cell contains and expresses a vector comprising the DNA coding for said heterologous protein operably linked to the DNA coding for the signal peptide OmpA or a functional derivative thereof.

Surprisingly, the use of the signal peptide OmpA alone and/ or in combination with the N-terminal amino acids SEGN/SEGNSD (SEQ ID NO:2/SEQ ID NO:3) translocate the recombinant DNA-derived proteins to the outer surface and facilitates the release of the functional and active molecule into the culture medium to a greater extent than any other method in the prior art. Before crossing the outer membrane, the recombinant DNA-derived protein is correctly folded according to the method of the present invention. The signal peptide is cleaved off to produce a mature molecule. Surprisingly, the efficiency of signal peptide removal is very high and leads to correct folding of the recombinant DNA-derived protein. This method according to the invention, exemplified for the kringle 2 plus serine protease domain (K2S) of tissue plasminogen activator protein in example 1 is generally applicable to expression of several different proteins and polypeptides which do not require mammalian glycosylation in prokaryotic host cells.

The method according to the invention has advantages over methods known in the art- not only that it is a cheap production method due to the prokaryotic host cell used, surprisingly, a correctly folded molecule is secreted to the supernatant.

The skilled person can easily obtain the DNA sequence of a protein of interest to be expressed by the method according to the invention from suitable databases and clone it to be used in the method according to the invention.

Said signal peptide OmpA interacts with SecE and is delivered across the inner membrane by energy generated by SecA, which binds to Sec components (SecE-SecY). SecY forms a secretion pore to dispatch the recombinant DNA-derived protein according to the invention. The space between the outer membrane and inner membrane of Gram-negative bacteria, periplasm, has higher oxidative condition in comparison to the cytoplasmic space. This supports the formation of disulfide bonds and properly folding of the recombinant protein (e.g. K2S) in the periplasm to yield an active molecule. According to the present invention, the signal peptide will be cleaved off to produce a mature molecule. The complex of GspD secretin and GspS lipoprotein on the outer membrane serves as gate channel for secreting the recombinant protein according to the invention to the extracellular medium. This secretion process requires energy, which is generated in cytoplasm by GspE nucleotide-binding protein then transferred to the inner membrane protein (Gsp G-J, F and K-N). GspC transfers the energy to GspD by forming a cross-linker between a set of inner membrane protein (Gsp G-J, F and K-N) and GspD. Before crossing the outer membrane successfully, the recombinant protein is correctly folded.

Operably linked according to the invention means that the DNA encoding the heterologous protein (preferably comprising the nucleic acid encoding SEGN or SEGNSD at its N-terminal portion) is cloned in close proximity to the OmpA DNA into the vector in order to achieve expression of the OmpA-heterologous protein-fusion protein and to direct secretion outside the prokaryotic host cell. Typically, the majority of the heterologous protein is secreted and can then be purified by appropriate methods such as ammonium sulfate precipitation. The invention also includes the use of inducers such as IPTG or IPTG in combination with glycerol, the improvement of the incubation condition and harvesting period to maximize the amount of active protein.

The inventors surprisingly found that the OmpA signal peptide alone or operatively linked to the amino acids characterized by the sequence SEGN (SEQ ID NO:2) or SEGNSD (SEQ ID NO:3)

lead to secretion of the heterologous protein into the medium rather than accumulation in the periplasmic space.

In a preferred embodiment, said DNA encoding the OmpA signal peptide may be fused to a short peptide characterized by the amino acid sequence SEGN (SEQ ID NO:2) or the coding nucleic acid sequence TCTGAGGGAAAC (SEQ ID NO:4) and located in the N-terminal portion or at the N-terminal portion of the heterologous protein. Thus, preferably, said fusion protein comprises OmpA-SEGN-heterologous protein. Even more preferred, said amino acids characterized by SEGN (SEQ ID NO:2) may be carry a point mutation or may be substituted by a non-natural amino acid. Even more preferred, there may be an amino acid or non-amino acid spacer between OmpA and SEGN or SEGN and the heterologous protein.

In a preferred embodiment, said DNA encoding the OmpA signal peptide may be fused to a short peptide characterized by the amino acid sequence SEGNSD (SEQ ID NO:3) or the coding nucleic acid sequence TCTGAGGGAAACAGTGAC (SEQ ID NO:5) and located in the N-terminal portion or at the N-terminal portion of the heterologous protein. Thus, preferably, said fusion protein comprises OmpA-SEGNSD-heterologous protein. Even more preferred, said amino acids characterized by SEGNSD may be carry a point mutation or may be substituted by a non-natural amino acid. Even more preferred, there may be an amino acid or non-amino acid spacer between OmpA and SEGNSD or SEGNSD and the heterologous protein.

Thus, in a preferred method according to the invention said the prokaryotic cell contains and expresses a vector comprising the DNA coding for said heterologous protein operably linked to the DNA coding for the signal peptide OmpA which is operably linked to the nucleic acid molecule defined by the sequence TCTGAGGGAAACAGTGAC (SEQ ID NO:5) or a functional derivative thereof.

Such heterologous proteins include, but are not limited to insulin, insulin-like growth factor, hGH, tPA, cytokines, e.g. interleukines (IL) such as IL-1, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-11, IL-12, IL-13, IL-14, IL-15, IL-16, IL-17, IL-18, interferon (IFN) alpha, IFN beta, IFN gamma, IFN omega or IFN tau, tumor necrosisfactor (TNF) TNF alpha and TNF beta, TRAIL; G-CSF, GM-CSF, M-CSF, MCP-1 and VEGF.

The method according to the invention can be advantageously used for production of antibodies or fragments thereof. Such fragments include e.g. Fab fragments (Fragment antigen-binding = Fab). Fab fragments consist of the variable regions of both chains which are held together by the adjacent constant region. These may be formed by protease digestion, e.g. with papain, from

conventional antibodies, but similar Fab fragments may also be produced in the mean time by genetic engineering. Further antibody fragments include F(ab')₂ fragments, which may be prepared by proteolytic cleaving with pepsin.

Using genetic engineering methods it is possible to produce shortened antibody fragments which consist only of the variable regions of the heavy (VH) and of the light chain (VL). These are referred to as Fv fragments (Fragment variable = fragment of the variable part). Since these Fv-fragments lack the covalent bonding of the two chains by the cysteines of the constant chains, the Fv fragments are often stabilised. It is advantageous to link the variable regions of the heavy and of the light chain by a short peptide fragment, e.g. of 10 to 30 amino acids, preferably 15 amino acids. In this way a single peptide strand is obtained consisting of VH and VL, linked by a peptide linker. An antibody protein of this kind is known as a single-chain-Fv (scFv). Examples of scFv-antibody proteins of this kind known from the prior art are described in Huston et al. (1988, PNAS 16: 5879-5883).

In recent years, various strategies have been developed for preparing scFv as a multimeric derivative. This is intended to lead, in particular, to recombinant antibodies with improved pharmacokinetic and biodistribution properties as well as with increased binding avidity. In order to achieve multimerisation of the scFv, scFv were prepared as fusion proteins with multimerisation domains. The multimerisation domains may be, e.g. the CH₃ region of an IgG or *coiled coil* structure (helix structures) such as *Leucine-zipper* domains. However, there are also strategies in which the interaction between the VH/VL regions of the scFv are used for the multimerisation (e.g. dia-, tri- and pentabodies). By diabody the skilled person means a bivalent homodimeric scFv derivative (Hu et al., 1996, PNAS 16: 5879-5883). The shortening of the *Linker* in an scFv molecule to 5- 10 amino acids leads to the formation of homodimers in which an inter-chain VH/VL-superimposition takes place. Diabodies may additionally be stabilised by the incorporation of disulphide bridges. Examples of diabody-antibody proteins from the prior art can be found in Perisic et al. (1994, Structure 2: 1217-1226).

By minibody the skilled person means a bivalent, homodimeric scFv derivative. It consists of a fusion protein which contains the CH₃ region of an immunoglobulin, preferably IgG, most preferably IgG1 as the dimerisation region which is connected to the scFv via a *Hinge region* (e.g. also from IgG1) and a *Linker* region. The disulphide bridges in the *Hinge* region are mostly formed in higher cells and not in prokaryotes. Examples of minibody-antibody proteins from the prior art can be found in Hu et al. (1996, Cancer Res. 56: 3055-61).

By triabody the skilled person means a: trivalent homotrimeric scFv derivative (Kortt et al. 1997 Protein Engineering 10: 423-433). ScFv derivatives wherein VH-VL are fused directly without a linker sequence lead to the formation of trimers.

The skilled person will also be familiar with so-called miniantibodies which have a bi-, tri- or tetraivalent structure and are derived from scFv. The multimerisation is carried out by di-, tri- or tetrameric coiled coil structures (Pack et al., 1993 Biotechnology 11:, 1271-1277; Lovejoy et al. 1993 Science 259: 1288-1293; Pack et al., 1995 J. Mol. Biol. 246: 28-34).

Therefore in another preferred method according to the invention an antibody or antibody fragment as described supra is produced.

The method according to the invention comprises prokaryotic host cells such as, but not limited to *Escherichia coli* (*E. coli*), *Bacillus subtilis*, *Streptomyces*, *Pseudomonas*, e.g. *Pseudomonas putida*, *Proteus mirabilis* or *Staphylococcus*, e.g. *Staphylococcus carnosus*. Preferably said host cells according to the invention are Gram-negative bacteria.

Preferably, a method according to the invention is also characterised in that the prokaryotic cell is *E. coli*. Suitable strains include, but are not limited to *E.coli* XL-1 blue, *E.coli* BL21(DE3), *E.coli* JM109, *E.coli* DH series, *E.coli* TOP10 and *E.coli* HB101.

Preferably, a method according to the invention is also characterised in that the following steps are carried out:

- a) the DNA encoding the heterologous protein is amplified by PCR;
- b) the PCR product is purified;
- c) said PCR product is inserted into a vector comprising the DNA coding for OmpA signal peptide and the DNA coding for gpIII in such a way that said PCR product is operably linked upstream to the DNA coding for the OmpA signal sequence and linked downstream to the DNA coding for gpIII of said vector;
- d) that a stop codon is inserted between said heterologous protein and gpIII;
- e) said vector is expressed by the prokaryotic cell
- f) the heterologous protein is purified.

For step a) according to the invention the choice / design of the primers is important to clone the DNA in the right location and direction of the expression vector (see example 1). Thus, the primers as exemplified in example 1 and figure 4 comprise an important aspect of the present invention. With gp III of step c) gene protein III is meant which is present mainly in phagemid vectors. The stop codon is inserted to avoid transcription of gp III thus eventually leading to

secretion of the heterologous protein of interest. Any suitable method for insertion of the stop codon may be employed such as site-directed mutagenesis (e.g. Weiner MP, Costa GL (1994) PCR Methods Appl 4(3):S131-136; Weiner MP, Costa GL, Schoettlin W, Cline J, Mathur E, Bauer JC (1994) Gene 151(1-2):119-123; see also example 1).

Any vector may be used in the method according to the invention, preferably said vector is a phagemid vector (see below).

The untranslated region may contain a regulatory element, such as e.g. a transcription initiation unit (promoter) or enhancer. Said promoter may, for example, be a constitutive, inducible or development-controlled promoter. Preferably, without ruling out other known promoters, the constitutive promoters of the human Cytomegalovirus (CMV) and Rous sarcoma virus (RSV), as well as the Simian virus 40 (SV40) and Herpes simplex promoter. Inducible promoters according to the invention comprise antibiotic-resistant promoters, heat-shock promoters, hormone-inducible, „Mammary tumour virus promoter“ and the metallothioneine promoter. Preferred promoters include T3 promoter, T7 promoter, Lac/ara1 and Ltet0-1.

More preferably, a method according to the invention is also characterised in that the DNA encoding the heterologous protein is preceded by a lac promoter and/or a ribosomal binding site such as the Shine-Dalgarno sequence (see also example).

Suitable vectors according to the invention include, but are not limited to viral vectors such as e.g. Vaccinia, Semliki-Forest-Virus and Adenovirus, phagemid vectors and the like. Preferred are vectors which can be advantageously used in E. coli, but also in any other prokaryotic host such as pPROTet.E, pPROLar.A, members of the pBAD family, pSE family, pQE family and pCAL.

Another preferred embodiment of the invention relates to the vector pComb3HSS containing a DNA according to the invention, wherein the expression of the gp III protein is suppressed or inhibited by deleting the DNA molecule encoding said gp III protein or by a stop codon between the gene coding for a polypeptide containing the heterologous protein and the protein III gene.

Preferably, a method according to the invention is also characterised in that the heterologous protein is selected from human tissue plasminogen activator (tPA) or a fragment, a functional variant, an allelic variant, a subunit, a chemical derivative, a fusion protein or a glycosylation variant thereof. Such fragments, allelic variants, functional variants, variants based on the degenerative nucleic acid code, fusion proteins with an tPA protein according to the invention, chemical derivatives or a glycosylation variant of the tPA proteins according to the invention may include one, several or all of the following domains or subunits or variants thereof:

1. Finger domain (4-50)
2. Growth factor domain (50-87)
3. Kringle 1 domain (87-176)
4. Kringle 2 domain (176-262)
5. Protease domain (276-527)

The numbering/naming of the domains is according to Genbank accession number GI 137119 or Nature 301 (5897), 214-221 (1983).

More preferably, a method according to the invention is also characterised in that the heterologous protein is selected from the Kringle 2 (4.) plus Serine protease (5.) K2S variant of human tissue plasminogen activator or a fragment, a functional variant, an allelic variant, a subunit, a chemical derivative, a fusion protein or a glycosylation variant thereof.

More preferably, a method according to the invention is also characterised in that the vector is a phagemid vector comprising the DNA coding for OmpA signal peptide and the DNA coding for gpIII.

The following example is intended to aid the understanding of the invention and should in no way be regarded as limiting the scope of the invention.

Example 1

MATERIALS AND METHODS

Primer design. In order to amplify a specific part of tPA gene, a pair of primers sK2/174 [5' GAGGAGGAGGTGGCCAGGCGGCCTCTGAGGGAAACAGTGAC 3'] (SEQ ID NO:6) and ASSP [5' GAGGAGGAGCTGGCCGGCCTGGCCCGGTCGCATGTTGTCACG 3'] (SEQ ID NO:7) were synthesized (Life Technologies, Grand Island, NY). These primers were designed based on the human tPA gene retrieved from NCBI databases (g137119). They were synthesized with Sfi I end cloning sites (underlined) in such a way that the reading frame from the ATG of the gpIII gene in phagemid vector, pComb3HSS, will be maintained throughout the inserted sequence.

Another primer set for site-directed mutagenesis was designed to anneal at the sequence situated between K2S gene and gene III in pComb3H-K2S. The sequence of primers with mutation bases (underlined) for generating a new stop codon were MS₁TPA [5'

10

ACATGCGACCGTGACAGGCCGGCCAG 3'] (SEQ ID NO:8) and MAS^{TPA} [5'
CTGGCCGGCCTGTCACGGTCGCATGT 3'] (SEQ ID NO:9).

Amplification of K2S gene by PCR. One μ g sK2/174 and AS^{SP} primers together with 50
ng of p51-3 template (obtained from Dr. Hiroshi Sasaki, Fujisawa Pharmaceutical, Japan) were
suspended in 100 μ l PCR mixture. An amount of 2.5 U Taq polymerase (Roche Molecular
Biochemicals, Indianapolis, IN) was finally added to the solution. The titrated amplification
condition was initiated with jump start at 85°C for 4 min, then denaturation at 95°C for 50 sec,
annealing at 42°C for 50 sec, extension at 72°C for 1.5 min. Thirty five rounds were repeatedly
performed. The mixture was further incubated at 72°C for 10 min. The amplified product of 1110
bp was subsequently purified by QIAquick PCR Purification Kit (QIAGEN, Hilden, Germany).
The correctness of purified product was confirmed by restriction enzymes.

Construction of phagemid expressing K2S. The purified PCR product of K2S and
pComb3HSS phagemid (kindly provided by Dr. Carlos F. Barbas, Scripps Institute, USA) were
digested with Sfi I (Roche Molecular Biochemicals, Indianapolis, IN) to prepare specific
cohesive cloning sites. Four μ g of the purified PCR product was digested with 60 U of Sfi I at
50°C for 18 h. For pComb3HSS, 20 μ g of phagemid vectors were treated with 100 U of Sfi I.
Digested products of purified PCR product of K2S and pComb3HSS (~3300 bp) were
subsequently gel-purified by the QIAquick Gel Extraction Kit (QIAGEN, Hilden, Germany). T4
ligase (Roche Molecular Biochemicals, Indianapolis, IN) of 5 U were introduced to the mixture
of 0.7 μ g of purified Sfi I-digested pComb3HSS and 0.9 μ g of purified Sfi I-digested PCR
product. Ligation reaction was incubated at 30°C for 18 h. The newly constructed phagemid was
named pComb3H-K2S.

Transformation of XL-1 Blue. Two hundred μ l of CaCl₂ competent *E. coli* XL-1 Blue
(Stratagene, La Jolla, CA) were transformed with 70 ng of ligated or mutated product. The
transformed cells were propagated by spreading on LB agar containing 100 μ g/ml ampicillin and
10 μ g/ml tetracycline (Sigma, Saint Louis, MO). After cultivation at 37°C for 18 h several
antibiotic resistant colonies were selected for plasmid minipreps by using the alkaline lysis
method. Each purified plasmid was subjected to Sfi I restriction site analysis. A transformant

harboring plasmid with the correct Sfi I restriction site(s) was subsequently propagated for 18 h at 37°C in 100 ml LB broth with ampicillin 100 µg/ml and tetracycline 10 µg/ml. A plasmid maxiprep was performed using the QIAGEN Plasmid Maxi Kit (QIAGEN, Hilden, Germany). The purified plasmid was reexamined for specific restriction sites by Sfi I and sequenced by AmpliTaq DNA Polymerase Terminator Cycle Sequencing Kit (The Perkin-Elmer Corporation, Forster City, CA).

Site-directed mutagenesis of pComb3H-K2S. 10 ng of pComb3H-K2S template were mixed with 125 ng of _{MS}TPA and _{MAS}TPA primers. PfuTurbo DNA polymerase (Stratagene, LA Jolla, CA) of 2.5 U was added to the mixture for cycle amplification. The reaction started with one round of 95°C for 30 sec. Then it was followed by 16 rounds consisting of 95°C for 30 sec, 55°C for 1 min, and 68°C for 9 min. The reaction tube was subsequently placed on ice for 2 min. In order to destroy the template strands, 10 U of Dpn I restriction enzyme (Stratagene, LA Jolla, CA) were added to the amplification reaction and incubated for 1 h at 37°C. This synthesized product (MpComb3H-K2S) was further used to transform *E. coli* XL-1 Blue.

Preparation of phage-display recombinant-K2S. After pComb3H-K2S was transformed to *E. coli* XL-1 Blue, the phage display technique was performed. A clone of pComb3H-K2S transformed *E. coli* XL-1 Blue was propagated in 10 ml super broth containing ampicillin 100 µg/ml and tetracycline 10 µg/ml at 37°C until the O.D. [600 nm] of 1.5 was reached. The bacterial culture was subsequently propagated in 100 ml of the same medium and culture for 2 h. An amount of 10¹² pfu of VCSM13 helper phage (Stratagene, La Jolla, CA) was used to infect the transformed *E. coli* XL-1 Blue. After 3 h incubation, kanamycin at a final concentration of 70 µg/ml final concentration was added to culture. The culture was left shaking (200 RPM) for 18 h at 37°C. Bacteriophages which harbored K2S on gp3 (K2S-φ) were then harvested by adding 4% w/v PEG MW 8000 (Sigma, Saint Louis, MO) and 3% w/v NaCl. Finally, the harvested phage was resuspended in 2 ml PBS pH 7.4. The phage number was determined by infecting *E. coli* XL-1 Blue. The colony-forming unit per milliliter (cfu/ml) was calculated as described previously (21).

Expression of recombinant-K2S in shaker flasks. MpComb3H-K2S transformed XL-1 Blue was cultivated in 100 ml super broth (3% w/v tryptone, 2% w/v yeast extract and 1% w/v MOPS) at pH 7.0 in the presence of ampicillin (100 μ g/ml) at 37°C until an O.D. [600 nm] of 0.8 was reached. Subsequently, the protein synthesis was induced by 1 mM of IPTG (Promega, Madison, WI). The bacteria were further cultured shaking (200 RPM) for 6 h at 30°C. The culture supernatant was collected and precipitated with 55% saturated ammonium sulfate (32). The precipitate was reconstituted with PBS, pH 7.2, and dialysed in the same buffer solution at 4°C for 18 h. Periplasmic proteins from bacterial cells were extracted by using a chloroform shock as previously described by Ames et al. (2).

Immunoassay quantification of recombinant-K2S. In order to detect r-K2S, solid phase was coated with monoclonal anti-kringle 2 domain (16/B) (generously provided by Dr. Ute Zacharias, Central Institute of Molecular Biology, Berlin-Buch, Germany). The standard ELISA washing and blocking processes were preformed. Fifty μ l of 10^{11} cfu/ml of K2S- ϕ or secretory r-K2S were added into each anti-kringle 2 coated well. Antigen-antibody detection was carried out as follows. Either sheep anti-M13 conjugated HRP (Pharmacia Biotech, Uppsala, Sweden) or sheep anti-tPA conjugated HRP (Cedarlane, Ontario, Canada), was added to each reaction well after the washing step. The substrate TMB was subjected to every well and the reaction was finally ceased with H_2SO_4 solution after 30 min incubation. The standard melanoma tPA 86/670 (National Institute for Biological Standards and Control, Hertfordshire, UK) was used as positive control.

Amidolytic activity assay. A test kit for the detection of tPA amidolytic activity was purchased from Chromogenix (Molndal, Sweden). The substrate mixture containing plasminogen and S-2251 was used to determine serine protease enzymatic activity. The dilution of 10^{-2} of each ammonium precipitated sample was assayed with and without stimulator, human fibrinogen fragments. The assay procedure was according to the COASET t-PA manual.

SDS-PAGE and immunoblotting. The dialysed precipitate-product from culture supernatant was further concentrated 10 folds with centricon 10 (AMICON, Beverly, MA). The concentrated sample was subjected to protein separation by SDS-PAGE, 15% resolving gel, in the reducing buffer followed by electroblotting to nitrocellulose. The nitrocellulose was then

blocked with 4% skimmed milk for 2 hr. In order to detect r-K2S, a proper dilution of sheep anti-tPA conjugated HRP was applied to the nitrocellulose. The immunoreactive band was visualized by a sensitive detection system, Amplified Opti-4CN kit (BIORAD, Hercules, CA).

Copolymerized plasminogen polyacrylamide gel electrophoresis. An 11% resolving polyacrylamide gel was copolymerized with plasminogen and gelatin as previously described by Heussen et al. (14). The stacking gel was prepared as 4 % concentration without plasminogen and gelatin. Electrophoresis was performed at 4°C at a constant current of 8 mA. The residual SDS in gel slab was removed after gentle shaking at room temperature for 1h in 2.5% Triton X-100. Then the gel slab was incubated in 0.1 M glycine-NaOH, pH 8.3, for 5 h at 37°C. Finally, the gel slab was stained and destained by standard Coomassie brilliant blue (R-250) dying system. The location of the peptide harboring enzymatic activity was not stained by dye in contrast to blue-paint background.

RESULTS

Construction of K2S gene carrying vector. From the vector p51-3 we amplified the kringle 2 plus the serine protease portion of tPA (Ser¹⁷⁴ in kringle 2 domain to Pro⁵²⁷ in the serine protease) using primers _SK2/174 and _{AS}SP. The amplified 1110 bp product was demonstrated by agarose gel electrophoresis (Fig. 1, lane 2) and was inserted into pComb3HSS phagemid by double Sfi I cleavage sites on 5' and 3' ends in the correct reading frame. Thus a new vector, pComb3H-K2S, harboring the K2S was generated. In this vector K2S is flanked upstream by the OmpA signal sequence and downstream by gp3. The correct insertion of K2S was verified both by restriction analysis with Sfi I (Fig. 2, lane 3), PCR-analysis (demonstration of a single band at 1110 bp), and DNA sequencing. The schematic diagram of pComb3H-K2S map is given in Fig. 3.

Phage-displayed r-K2S. VCSM13 filamentous phage was used to infect pComb3H-K2S transformed *E. coli* XL-1 Blue, X[K2S]. VCSM13 was propagated and incorporated the K2S-gp3 fusion protein during the viral packaging processes. The harvested recombinant phage (K2S- ϕ) gave a concentration of 5.4×10^{11} cfu/ml determined by reinfecting *E. coli* XL-1 Blue with PEG-precipitated phages. These recombinant phage particles were verified for the expression of r-K2S by sandwich ELISA. The phage-bound heterologous K2S protein was recognized by the monoclonal anti-kringle 2 antibody (16/B) by using sheep anti-tPA conjugated HRP antibody

detection system. The absorbance of this assay was 1.12 ± 0.03 (Table 1). The amount of K2S detectable on 10^{12} phage particles is equal to 336 ng of protein in relation to the standard melanoma tPA. In order to corroborate that K2S-gp3 fusion protein was associated with phage particles, sheep anti-tPA conjugated HRP antibody was substituted by sheep anti-M13 antibody conjugated HRP. This immuno-reaction exhibited an absorbance of 1.89 ± 0.07 (Table 1). In contrast, if the capture antibody was sheep anti-M13 antibody, extremely low K2S was observed with sheep anti-tPA antibody conjugated HRP; the absorbance was only 0.17 ± 0.01 (Table 1). This suggested that only a minority of purified phage particles carried K2S-gp3 fusion protein. VCSM13 prepared from non-transformed *E. coli* XL-1 Blue was used as a negative control.

Construction of MpComb3H-K2S. We generated a stop codon between K2S and gp3 in pComb3H-K2S with the aid of the mutagenic primers ($M_{S}TPA$ and $M_{AS}TPA$) (Fig. 4). In order to enrich the newly synthesized and mutated MpComb3H-K2S, the cycle amplification mixture was thoroughly digested with Dpn I to degrade the old dam methylated pComb3H-K2S template (Dpn I prefers dam methylated DNA). After transforming of *E. coli* XL-1 Blue with MpComb3H-K2S, a transformant XM[K2S] was selected for further study. As a consequence of bp substitution, one Sfi I cleavage site close to the 3' end of K2S gene was lost after site-directed mutagenesis. A linear version of Sfi I cleaved MpComb3H-K2S was observed at 4319 bp without the appearance of inserted K2S gene fragment (Fig. 5, lane 3). Thus, the K2S gene encoding by MpComb3H-K2S was expressed in non-gp3 fusion form in *E. coli* XM[K2S].

Expression and purification of K2S. K2S expression in *E. coli* XM[K2S] was induced by IPTG. r-K2S was detectable by using ELISA both in the periplasmic space and in the culture supernatant. The amount of the heterologous protein in each preparation was determined by sandwich ELISA and related to the standard tPA. From 100 ml of the bacterial culture in shaker flask with the O.D. [600 nm] of 50, the periplasmic fraction yielded 1.38 μ g of r-K2S (approximately 32%) whereas 2.96 μ g of r-K2S (approximately 68%) was obtained in the ammonium precipitated culture supernatant. Sandwich ELISA was used to verify the PEG precipitated phage from VCSM13 infected *E. coli* XM[K2S]. No r-K2S captured by monoclonal anti-kringle 2 antibody was detected by anti-M13 conjugated HRP, indicating that K2S is not presented on the phage particles if gp3 is missing.

Amidolytic activity measurement. If serine protease domain is present in the sample, plasminogen will be converted to plasmin. The produced plasmin will further digest the S-2251 substrate to a colour product, p-nitroaniline, which has a maximum absorbance at 405 nm. The specific activity of the recombinant product is in accord with the absorbance. The fibrinogen-dependent enzymatic activity of each sample i.e. K2S- ϕ , periplasmic r-K2S or culture supernatant r-K2S, was evaluated and compared. Both K2S- ϕ and periplasmic r-K2S illustrated notably low enzymatic activity, which was below the sensitivity of the test (0.25 IU/ml). The culture supernatant r-K2S gave the fibrinogen-dependent enzymatic activity of 7 IU/ml. Thus, from 100 ml culture we obtained a total of 700 IU enzymatic activity. Without fibrinogen no enzymatic activity of the r-K2S purified from culture supernatant was observed - whereas standard melanoma tPA showed some activity.

Demonstration of recombinant protein by immunoblotting. Partially purified K2S from culture supernatant of *E. coli* XM[K2S] revealed a molecular mass of 39 kDa by using sheep anti-tPA antibodies (Fig. 6). The negative control, partially purified culture supernatant of non-transformed *E. coli* XL1-Blue, contained no reactive band with a similar size.

Localization of active enzyme by PAGE. The plasminogen has been copolymerized and immobilized with gelatin in the polyacrylamide gel prior to electrophoresis. The ammonium sulfate precipitated culture supernatants of *E. coli* XL-1 Blue, *E. coli* XL-1 Blue transformed with pComb3HSS and *E. coli* XM[K2S] were analyzed (Fig. 7). All samples were processed in non-reducing condition to preserve the correct conformation and activity of the serine protease domain. Transparent areas of serine protease digested plasminogen were observed only in the ammonium sulfate precipitated culture supernatants of *E. coli* XM[K2S] at 34 and 37 kDa positions. The other samples gave no clearing zones. The positive control lane of standard melanoma tPA also demonstrated enzymatic activity at 66 and 72 kDa positions.

References

1. Allen, S., H. Y. Naim, and N. J. Bulleid. 1995. Intracellular folding of tissue-type plasminogen activator. Effects of disulfide bond formation on N-linked glycosylation and secretion. *J. Biol. Chem.* 270:4797-4804.

2. Ames, G. F., C. Prody, and S. Kustu. 1984. Simple, rapid, and quantitative release of periplasmic proteins by chloroform. *J. Bacteriol.* 160:1181-1183.
3. Barbas, C. F. III, A. S. Kang, R. A. Lerner, and S. J. Benkovic. 1991. Assembly of combinatorial antibody libraries on phage surfaces: the gene III site. *Proc. Natl. Acad. Sci. U. S. A.* 88:7978-7982.
4. Barbas, C. F. III, and J. Wagner. 1995. Synthetic human antibodies: selecting and evolving functional proteins. *A Companion to Methods in Enzymology* 8: 94-103.
5. Bennett, W. F., N. F. Paoni, B. A. Keyt, D. Botstein, A. J. Jones, L. Presta, F. M. Wurm, and M. J. Zoller. 1991. High resolution analysis of functional determinants on human tissue-type plasminogen activator. *J Biol Chem.* 266:5191-5201.
- 10 6. Betton, J. M., N. Sassoon, M. Hofnung, and M. Laurent. 1998. Degradation versus aggregation of misfolded maltose-binding protein in the periplasm of *Escherichia coli*. *J. Biol. Chem.* 273:8897-8902.
7. Camiolo, S. M., S. Thorsen, and T. Astrup. 1971. Fibrinogenolysis and fibrinolysis with tissue plasminogen activator, urokinase, streptokinase-activated human globulin and plasmin. *Proc. Soc. Exp. Biol. Med.* 38:277-280.
- 15 8. Cartwright, T. 1992. Production of t-PA from animal cell culture, p. 217-245. In R. E. Spier, and J. B. Griffiths (ed.), *Animal Cell Biotechnology*, Vol 5. Academic Press, N.Y.
9. Curry, K. A., A. W. Yem, M. R. Deibel, Jr., N. T. Hatzenbuehler, J. G. Hoogerheide, and C. S. Tomich. 1990. *Escherichia coli* expression and processing of human interleukin-1 beta fused to signal peptides. *DNA Cell Biol.* 9:167-175.
- 20 10. Datar, R. V., T. Cartwright, and C.-G. Rosen. 1993. Process economics of animal cell and bacterial fermentations: a case study analysis of tissue plasminogen activator. *Biotechnology* 11:349-357.
11. Deneffe, P., S. Kovarik, T. Ciora, N. Gosselet, J. C. Benichou, M. Latta, F. Guinet, A. Ryter, and J. F. Mayaux. 1989. Heterologous protein export in *Escherichia coli*: influence of bacterial signal peptides on the export of human interleukin 1 beta. *Gene* 85:499-510.
- 25 12. Griffiths, J. B., A. Electricwala. 1987. Production of tissue plasminogen activators from animal cells. *Adv. Biochem. Eng. Biotechnol.* 34:147-166.
13. Harris, T. J., T. Patel, F. A. Marston, S. Little, J. S. Emtage, G. Opdenakker, G. Volckaert, W. Rombauts, A. Billiau, and P. De Somer. 1986. Cloning of cDNA coding for human tissue-type plasminogen activator and its expression in *Escherichia coli*. *Mol. Biol. Med.* 3:279-292.
- 30 14. Heussen, C., and E. B. Dowdle. 1980. Electrophoretic analysis of plasminogen activators in polyacrylamide gels containing sodium dodecyl sulfate and copolymerized substrates. *Anal. Biochem.* 102:196-202.
15. Heussen, C., F. Joubert, and E. B. Dowdle. 1984. Purification of human tissue plasminogen activator with *Erythrina* trypsin inhibitor. *J. Biol. Chem.* 259:11635-11638.

16. Hu, C. K., U. Kohnert, O. Wilhelm, S. Fischer, and M. Llinas. 1994. Tissue-type plasminogen activator domain-deletion mutant BM 06.022: modular stability, inhibitor binding, and activation cleavage. *Biochemistry* 33:11760-11766.
17. Kipriyanov, S. M., G. Moldenhauer, and M. Little. 1997. High level production of soluble single chain antibodies in small-scale *Escherichia coli* cultures. *J. Immunol. Methods* 200:69-77.
18. Ko, J. H., D. K. Park, I. C. Kim, S. H. Lee, and S. M. Byun. 1995. High-level expression and secretion of streptokinase in *Escherichia coli*. *Biotechnol. Lett.* 17:1019-1024.
19. Kouzuma, Y., N. Yamasaki, and M. Kimura. 1997. The tissue-type plasminogen activator inhibitor ETIa from *Erythrina variegata*: structural basis for the inhibitory activity by cloning, expression, and mutagenesis of the cDNA encoding ETIa. *J. Biochem. (Tokyo)* 121:456-463.
20. Lasters, I., N. Van Herzele, H. R. Lijnen, D. Collen, and L. Jespers. 1997. Enzymatic properties of phage-displayed fragments of human plasminogen. *Eur. J. Biochem.* 244:946-952.
21. Lobel, L. I., P. Rausch, I. Trakht, S. Pollak, and J. W. Lustbader. 1997. Filamentous phage displaying the extracellular domain of the hLH/CG receptor bind hCG specifically. *Endocrinology*. 138:1232-1239.
22. Lubiniecki, A., R. Arathoon, G. Polastri, J. Thomas, M. Wiebe, R. Garnick, A. Jones, R. van Reis, and S. Builder. Selected strategies for manufacture and control of recombinant tissue plasminogen activator prepared from cell culture, p. 442-451. In R. E. Spier, J. B. Griffiths, J. Stephenne, and P. J. Crooy (ed.), *Advances in animal cell biology and technology for bioprocesses*. Butterworths, London.
23. Lucic, M. R., B. E. Forbes, S. E. Grosvenor, J. M. Carr, J. C. Wallace, and G. Forsberg. 1998. Secretion in *Escherichia coli* and phage-display of recombinant insulin-like growth factor binding protein-2. *J. Biotechnol.* 61:95-108.
24. Martin, U., S. Fischer, U. Kohnert, H. Lill, R. Rudolph, G. Sponer, A. Stern, and K. Strein. 1990. Properties of a novel plasminogen activator (BM 06.022) produced in *Escherichia coli*. *Z. Kardiol.* 79:167-170.
25. Obukowicz, M. G., M. E. Gustafson, K. D. Junger, R. M. Leimgruber, A. J. Wittwer, T. C. Wun, T. G. Warren, B. F. Bishop, K. J. Mathis, D. T. McPherson, N. R. Siegel, M. G. Jennings, B. B. Brightwell, J. A. Diaz-Cuellar, L. D. Bell, C. S. Craik, and W. C. Tacon. 1990. Secretion of active kringle-2-serine protease in *Escherichia coli*. *Biochemistry* 29:9737-9745.
26. Parmley, S. F., and G. P. Smith. 1988. Antibody-selectable filamentous fd phage vectors: affinity purification of target genes. *Gene* 73:305-318.
27. Pennica, D., W. E. Holmes, W. J. Kohr, R. N. Harkins, G. A. Vohar, C. A. Ward, W. F. Bennett, E. Yelverton, P. H. Seeburg, H. I. Heyneker, D. V. Goeddel, and D. Collen. 1983. Cloning and expression of human tissue-type plasminogen activator cDNA in *E. coli*. *Nature* 301:214-221.

28. Rippmann, J. F., M. Klein, C. Hoischen, B. Brocks, W. J. Rettig, J. Gumpert, K. Pfizenmaier, R. Mattes, and D. Moosmayer. 1998. Prokaryotic expression of single-chain variable-fragment (scFv) antibodies: secretion in L-form cells of *Proteus mirabilis* leads to active product and overcomes the limitations of periplasmic expression in *Escherichia coli*. *Appl. Environ. Microbiol.* 64:4862-4869.
29. Saito, Y., Y. Ishii, H. Sasaki, M. Hayashi, T. Fujimura, Y. Imai, S. Nakamura, S. Suzuki, J. Notani, T. Asada, H. Horiai, K. Masakazu, and N. Mineo. 1994. Production and characterization of a novel tissue-type plasminogen activator derivative in *Escherichia coli*. *Biotechnol. Prog.* 10:472-479.
30. Sarmientos, P., M. Duchesne, P. Denèfle, J. Boiziau, N. Fromage, N. Delporte, F. Parker, Y. Lelièvre, J.-F. Mayaux, and T. Cartwright. 1989. Synthesis and purification of active human tissue plasminogen activator from *Escherichia coli*. *Biotechnology* 7:495-501.
31. Scherrer, S., N. Robas, H. Zouheiry, G. Branlant, and C. Branlant. 1994. Periplasmic aggregation limits the proteolytic maturation of the *Escherichia coli* penicillin G amidase precursor polypeptide. *Appl. Microbiol. Biotechnol.* 42:85-89.
32. Soeda, S., M. Kakiki, H. Shimeno, and A. Nagamatsu. 1986. Rapid and high-yield purification of porcine heart tissue-type plasminogen activator by heparin-sepharose chromatography. *Life Sci.* 39:1317-1324.
33. Szarka, S. J., E. G. Sihota, H. R. Habibi., and S.-L. Wong. 1999. Staphylokinase as a plasminogen activator component in recombinant fusion proteins. *Appl. Environ. Microbiol.* 65:506-513.
34. Waldenström, M., E. Holmgren, A. Attersand, C. Kalderen, B. Lowenadler, B. Raden, L. Hansson, and G. Pohl. 1991. Synthesis and secretion of a fibrinolytically active tissue-type plasminogen activator variant in *Escherichia coli*. *Gene* 99:243-248.
35. Wan, E. W.-M., and F. Baneyx. 1998. TolAIII Co-overexpression Facilitates the Recovery of Periplasmic Recombinant Proteins into the Growth Medium of *Escherichia coli*. *Protein Expr. Purif.* 14:13-22.
36. Zacharias, U., B. Fischer, F. Noll, and H. Will. 1992. Characterization of human tissue-type plasminogen activator with monoclonal antibodies: mapping of epitopes and binding sites for fibrin and lysine. *Thromb. Haemost.* 67:88-94.

Figure Legends

- FIG. 1. Validation of PCR amplification product of the K2S gene from the p51-3 vector by using $s_{K2/174}$ and A_{SP} primers. Lane 1 shows 1 kb marker (Roche Molecular Biochemicals, Indianapolis, IN). Lane 2 was loaded with 1 μ l of amplified product. A single band at 1110 bp is depicted. The electrophoresis was performed on a 1% agarose gel.

FIG. 2. Identification of inserted K2S gene at 1110 bp () after Sfi I digested pComb3H-K2S was demonstrated in lane 3. Lane 1 shows 1 kb marker. Lane 2 was loaded with uncut pComb3H-K2S. The electrophoresis was performed on a 1% agarose gel.

FIG. 3. Scheme of pComb3H-K2S showing two Sfi I cloning sites into which the K2S gene was inserted. Signal sequence (OmpA), ribosome binding site (RIBS), lac promotor, and gpIII gene are also depicted.

FIG. 4. Schematic diagram of the mutation site at the junction between the K2S and gpIII genes on pComb3H-K2S. The annealing site of pComb3H-K2S is bound with a set of mutation primers ($M_{S}TPA$ and $M_{AS}TPA$) containing modified oligonucleosides (underlined). After performing the cycle amplification, the Sfi I site 1 (in bold) is modified and lost in the newly synthesized strand.

FIG. 5. Characterization of newly synthesized MpComb3H-K2S by the Sfi I restriction enzyme. A single band at 4319 bp that refers to a single cleavage site of MpComb3H-K2S is observed in lane 3. No inserted K2S band at 1110 bp can be visualized. Lane 1 shows 1 kb marker. Lane 2 was loaded with uncut MpComb3H-K2S. The electrophoresis was performed on a 1% agarose gel.

FIG. 6. Identification of immunological reactive band with of recombinant protein purified from *E. coli* XM[K2S] culture supernatant with sheep anti-tPA conjugated HRP. Lane 1 was loaded with 40 ng of standard melanoma tPA (86/670), which showed the reactive band at 70 kDa. The partially purified and concentrated culture supernatants from non-transformed *E. coli* XL1- Blue and *E. coli* XM[K2S] were applied to lane 2 and 3 respectively. The distinct reactive band was particularly demonstrated in lane 3 at 39 kDa.

FIG. 7. Molecular weight determination of extracellular r-K2S harboring active serine protease domain by copolymerized plasminogen polyacrylamide gel electrophoresis. Lane 1 contained the indicated molecular weight standards ($\times 10^{-3}$), SDS-6H (Sigma, Saint Louis, MO). Fifty μ g of the 55% saturated ammonium sulfate precipitated culture supernatant of *E. coli* XL-1 Blue, *E. coli* XL-1 Blue transformed with pComb3HSS, and *E. coli* XM[K2S] were loaded in lane 2, 3, and 4 respectively. Lane 5 contained 50 mIU of standard melanoma tPA (86/670). Transparent

zones of digested plasminogen in polyacrylamide gel are visible only in lane 4 at molecular weight of 34 and 37 kDa (B) and lane 5 at molecular weight of 66 and 72 kDa (A).

FIG. 8. Structure A

Native K2S molecule from amino acids 174-527 without modification (SEQ ID NO:10).

FIG. 9. Structure B-0

Native K2S molecule from amino acids 197-527 without modification. (SEQ ID NO:11)

FIG. 10. Structure B-1

K2S molecule from amino acids 193-527, wherein to Structure B-0 of Fig. 9 the amino acids SEGN were added at the N-terminal portion (SEQ ID NO:12).

FIG. 11. Structure B-2

K2S molecule from amino acids 193-527, as in Fig. 10, wherein Cys-261 was exchanged for Ser (SEQ ID NO:13).

FIG. 12. Structure B-3

K2S molecule from amino acids 191-527, wherein to Structure B-0 of Fig. 9 the amino acids SEGNSD were added at the N-terminal portion (SEQ ID NO:14).

FIG. 13. Structure B-4

K2S molecule from amino acids 191-527, as in Fig. 12, wherein Cys-261 was exchanged for Ser (SEQ ID NO:15).

FIG. 14. Structure C

Native K2S molecule from amino acids 220-527 without modification. This molecule may be further modified in a similar manner as disclosed for structure B in figures 10-13 (SEQ ID NO:16).

FIG. 15. Structure D

Native K2S molecule from amino acids 260-527 without modification. This molecule may be further modified in a similar manner as disclosed for structure B in figures 10-13 (SEQ ID NO:17).

FIG. 16. tPA molecule (SEQ ID NO:18)

Table

TABLE 1. Detection of r-K2S molecule in phage preparation by sandwich ELISA

Capture antibody	Tracer antibody (conjugated HRP)			
	Anti-tPA		Anti-M13	
	K2S- ϕ	VCSM13 ^a	K2S- ϕ	VCSM13
Anti-kringle 2 ^b	1.12 \pm 0.04 ^c	0.12 \pm 0.03	1.89 \pm 0.02	0.16 \pm 0.02
Anti-M13	0.17 \pm 0.01	0.14 \pm 0.05	1.91 \pm 0.02	1.88 \pm 0.03

^a VCSM13 was harvested from XL-1 Blue transformed with pComb3HSS.

^b Mouse monoclonal anti-kringle 2 (16/B) was used. The other antibodies were prepared from sheep immunoglobulin.

^c Value is mean of absorbance of each sample which was assayed in triplicate.

Claims

1. Method for the production of recombinant DNA-derived heterologous protein in prokaryotic cells, wherein said heterologous protein is secreted extracellularly as an active and correctly folded protein, characterized in that the prokaryotic cell contains and expresses a vector comprising the DNA coding for said heterologous protein operably linked to the DNA coding for the signal peptide OmpA or a functional derivative thereof.

2. Method according to claim 1, characterised in that said the prokaryotic cell contains and expresses a vector comprising the DNA coding for said heterologous protein operably linked to the DNA coding for the signal peptide OmpA which is operably linked to the nucleic acid molecule defined by the sequence TCTGAGGGAAACAGTGAC (SEQ ID NO:5) or a functional derivative thereof.

3. Method according to claim 1 or 2, characterised in that the prokaryotic cell is *E. coli*.

4. Method according to one of claims 1 to 3, characterised in that the the following steps are carried out:

- a) the DNA encoding the heterologous protein is amplified by PCR;
- b) the PCR product is purified;
- c) said PCR product is inserted into a vector comprising the DNA coding for OmpA signal peptide and the DNA coding for gpIII in such a way that said PCR product is operably linked upstream to the DNA coding for the OmpA signal sequence and linked downstream to the DNA coding for gpIII of said vector;
- d) that a stop codon is inserted between said heterologous protein and gpIII;
- e) said vector is expressed by the prokaryotic cell;
- f) the heterologous protein is purified.

5. Method according to one of claims 1 to 4, characterised in that the heterologous protein is selected from human tissue plasminogen activator or a fragment, a functional variant, an allelic variant, a subunit, a chemical derivative, a fusion protein or a glycosylation variant thereof.

6. Method according to one of claims 1 to 5, characterised in that the heterologous protein is selected from the K2S variant of human tissue plasminogen activator or a fragment, a functional

variant, an allelic variant, a subunit, a chemical derivative, a fusion protein or a glycosylation variant thereof.

7. Method according to one of claims 1 to 6, characterised in that the vector is a phagemid vector comprising the DNA coding for OmpA signal peptide and the DNA coding for gpIII.

8. Method according to one of claims 1 to 7, characterised in that the vector is the pComb3HSS phagemid.

9. Method according to one of claims 1 to 8, characterised in that the DNA Sequence of OmpA comprises the following sequence:

ATGAAAAAGACAGCTATCGCGATTGCAGTGGCACTGGCTGGTTTCGCTACCGTGGCC
CAGGCGGCC (SEQ ID NO:1)

10. Method according to one of claims 1 to 9, characterised in that the DNA Sequence of OmpA consists of the following sequence:

ATGAAAAAGACAGCTATCGCGATTGCAGTGGCACTGGCTGGTTTCGCTACCGTGGCC
CAGGCGGCC (SEQ ID NO:1)

11. Method according to one of claims 1 to 10, characterised in that the DNA of the heterologous protein is preceded by a lac promotor and/or a ribosomal binding site.

1/15

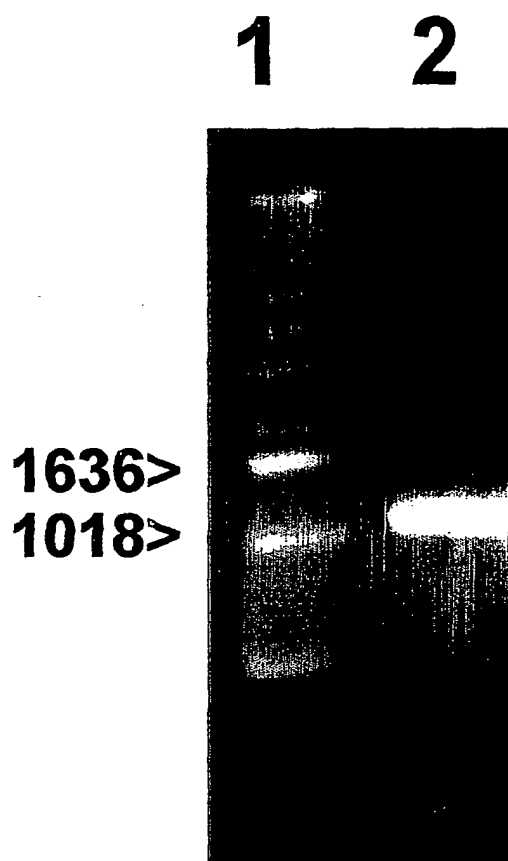
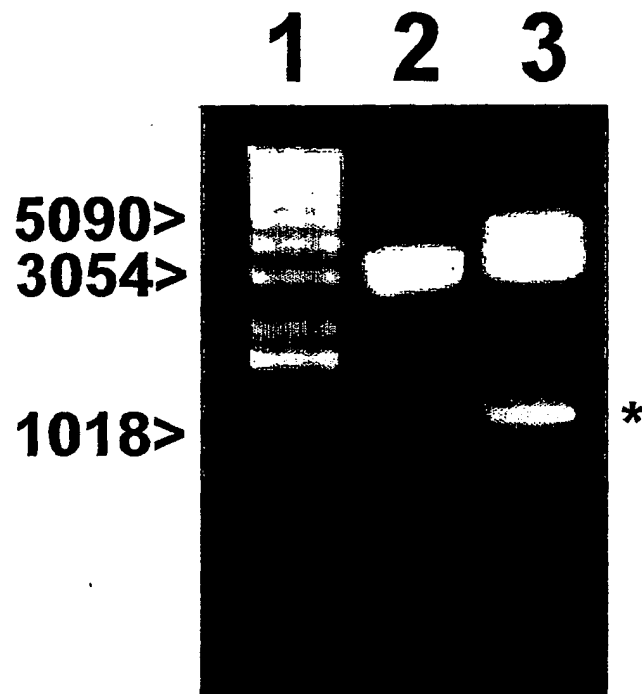


Fig 1

2/15

**Fig 2**

3/15

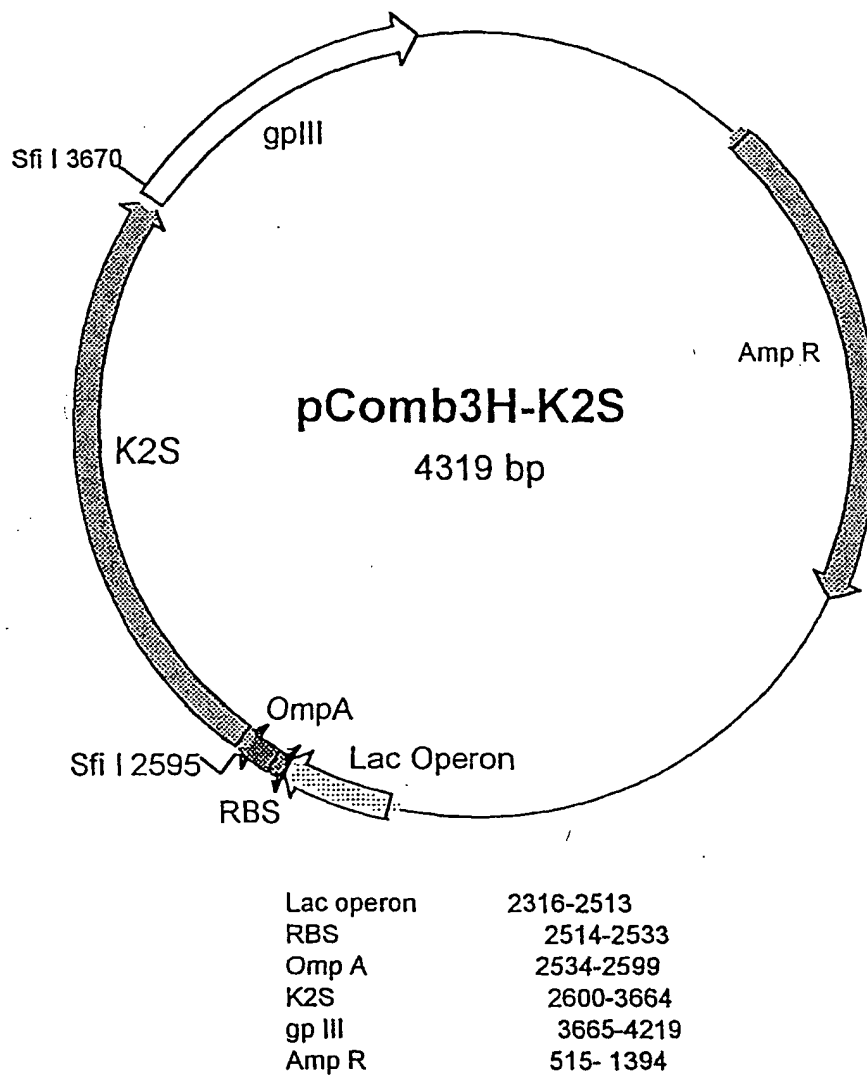


Figure 3

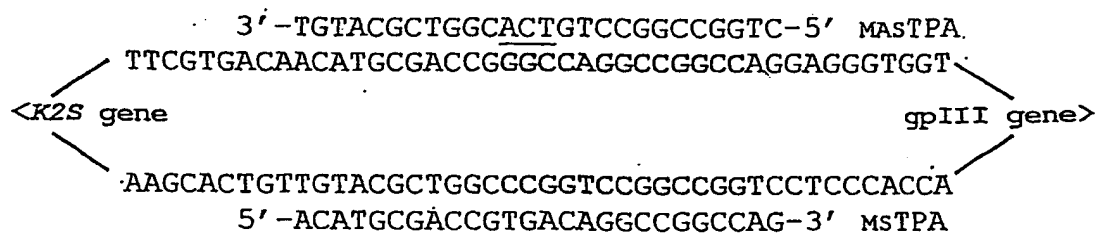
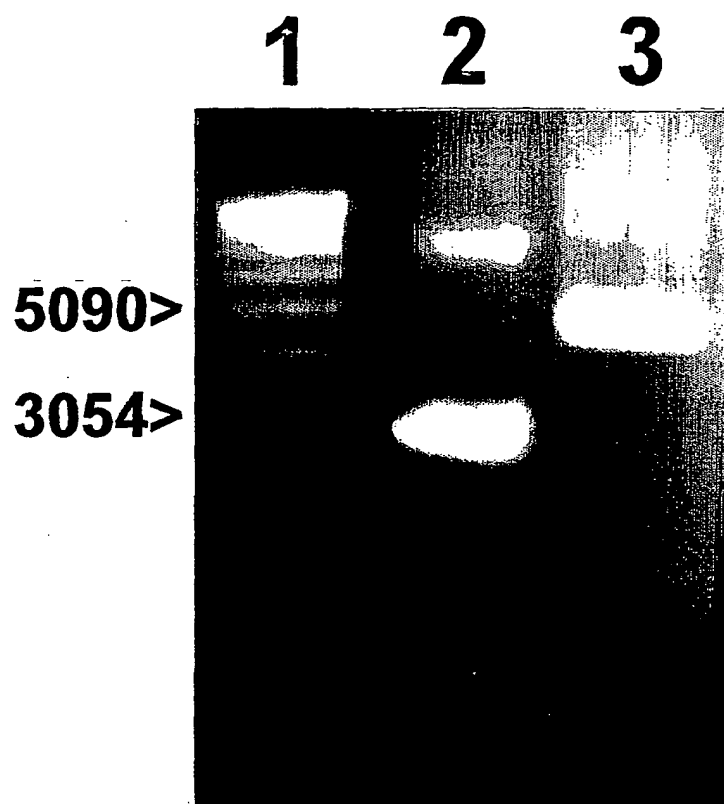
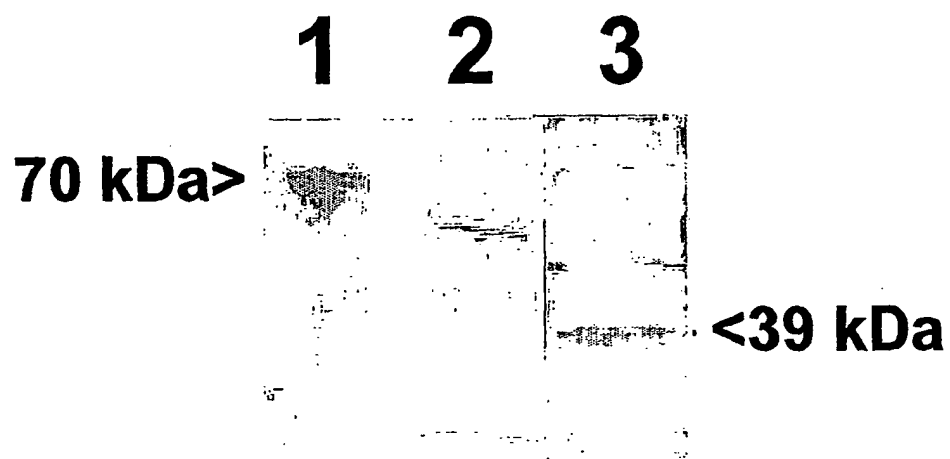


Figure 4

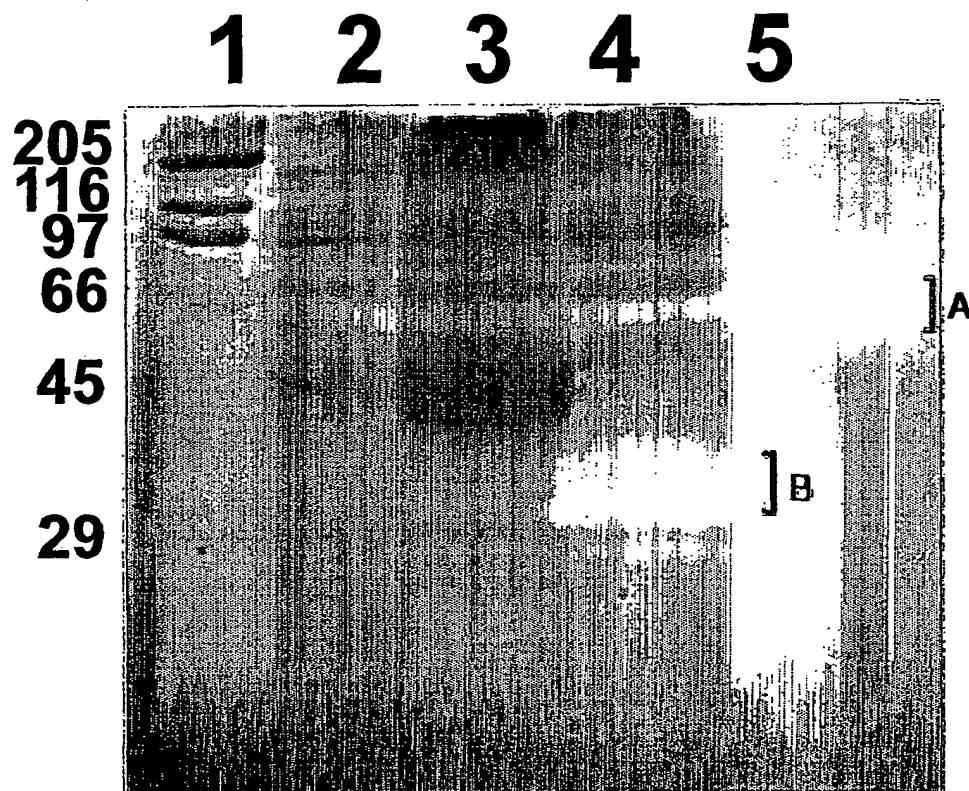
4/15

**Fig 5**

5/15

**Fig 6**

6/15

**Fig 7**

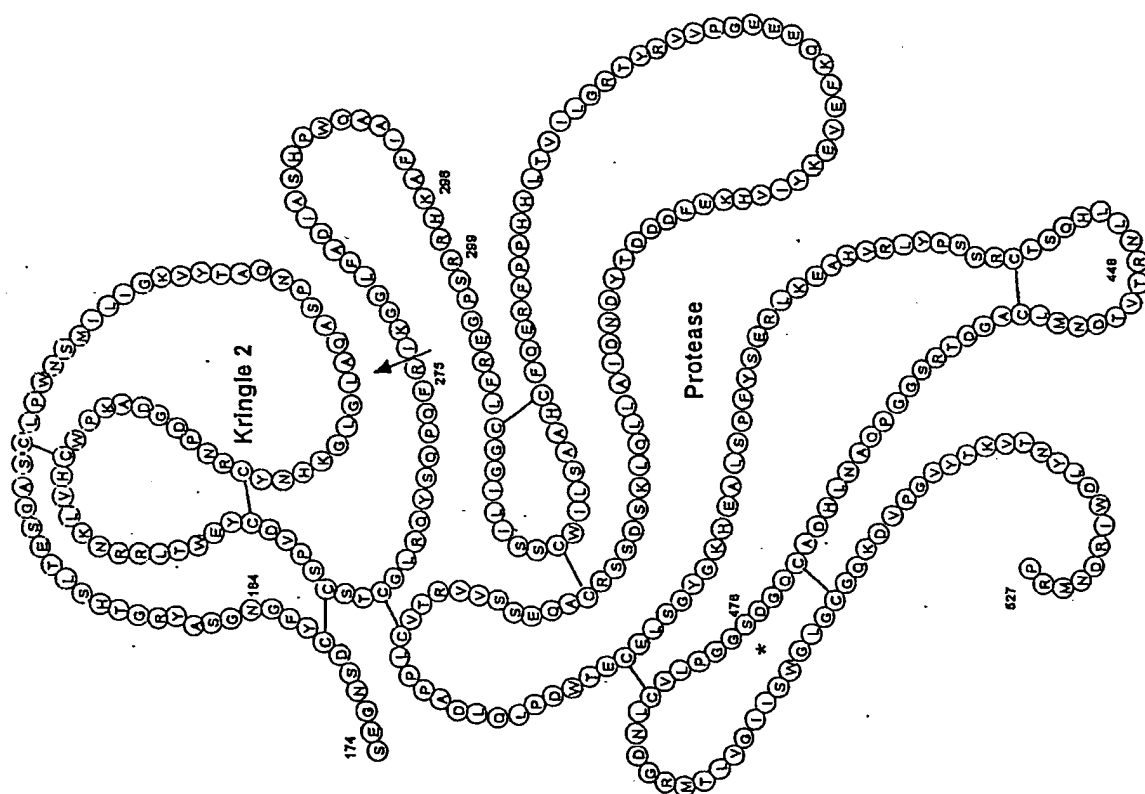
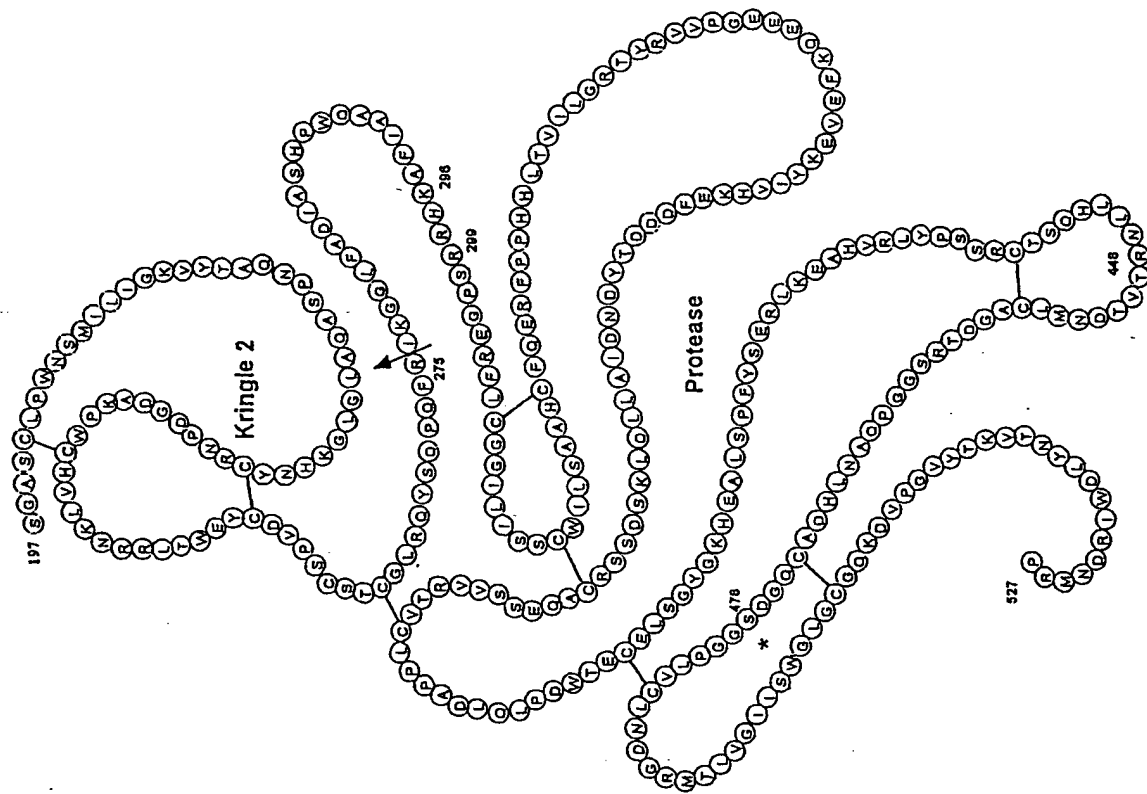


Fig. 8

8/15



9/15

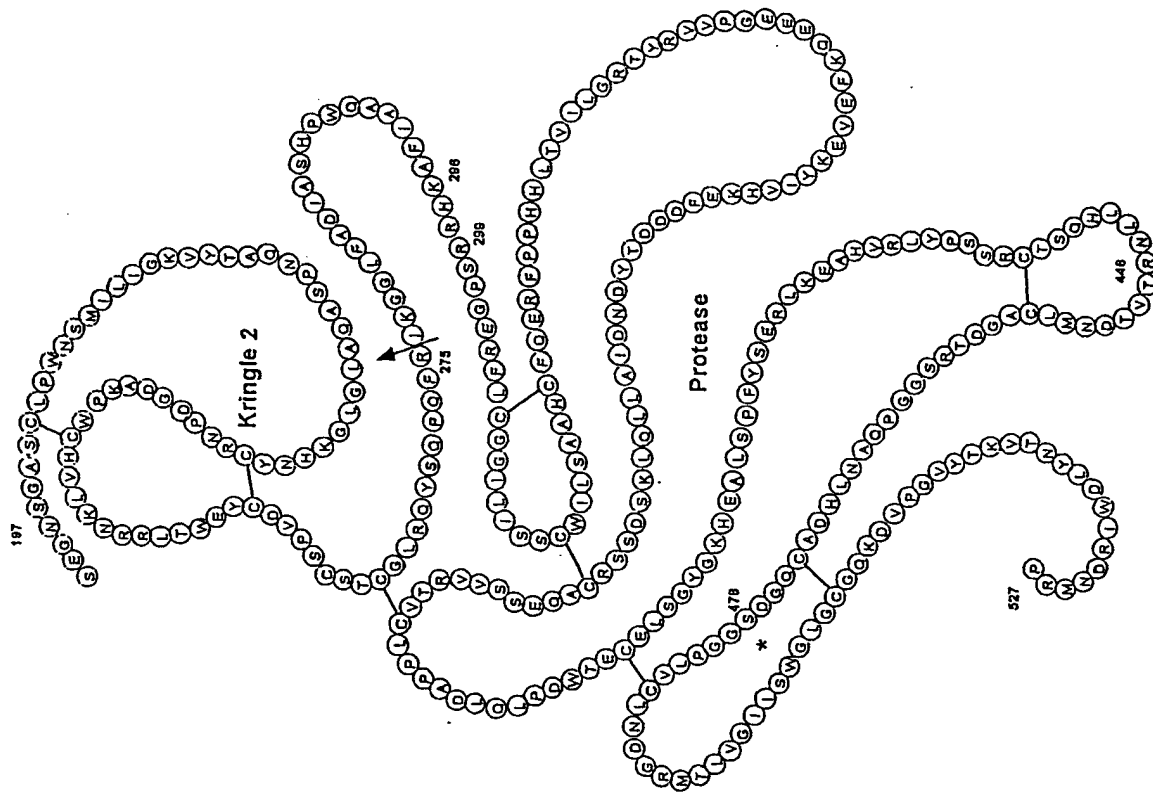


Fig. 10

10/15

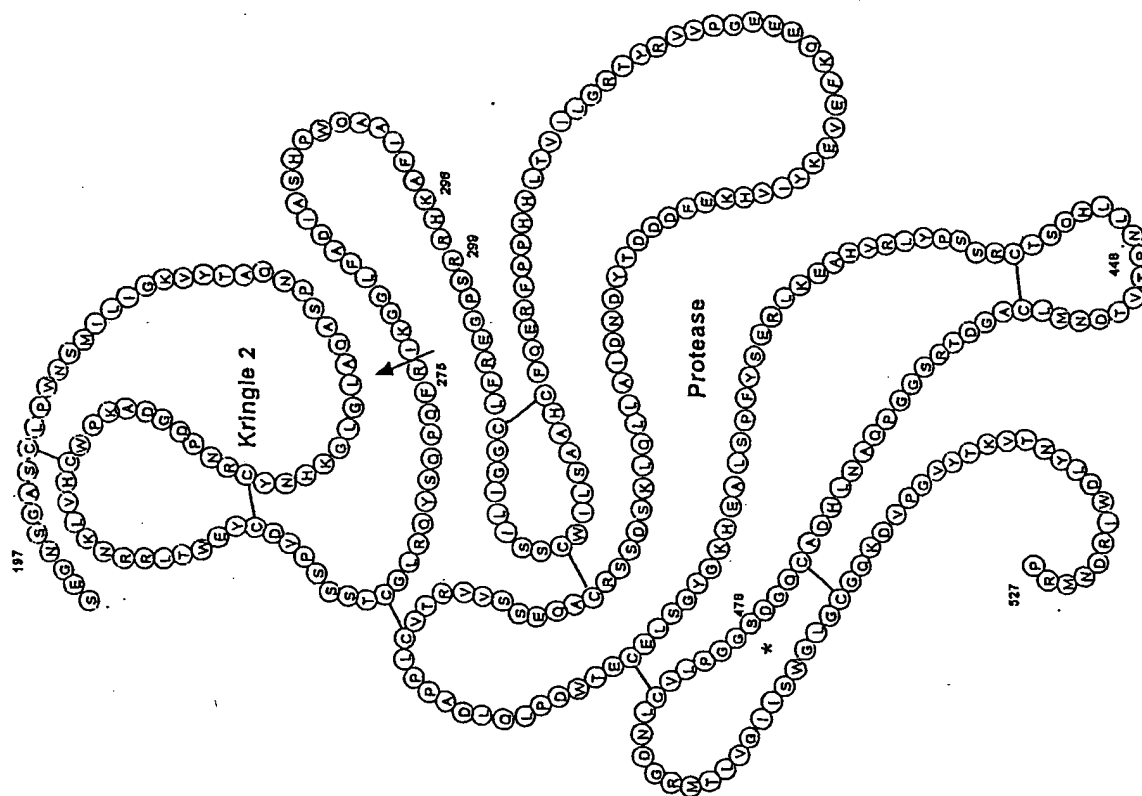


Fig. 11

11/15

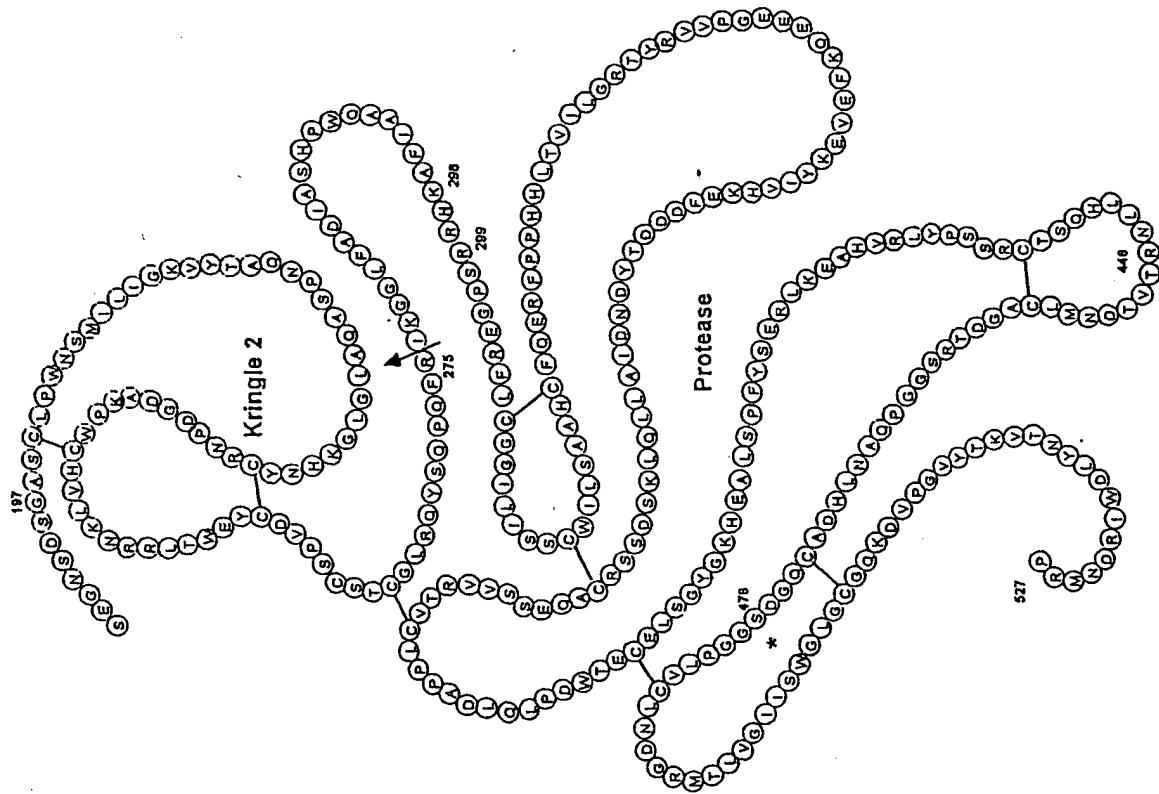


Fig. 12

12/15

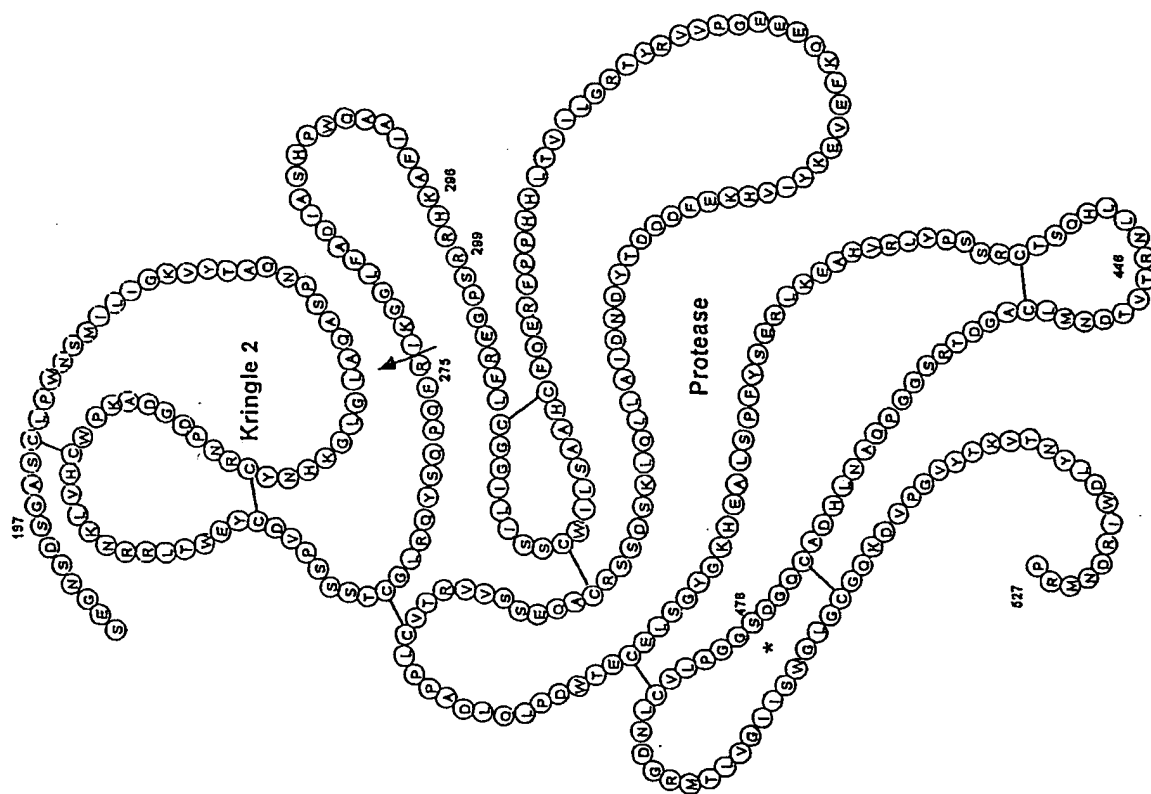


Fig. 13

13/15

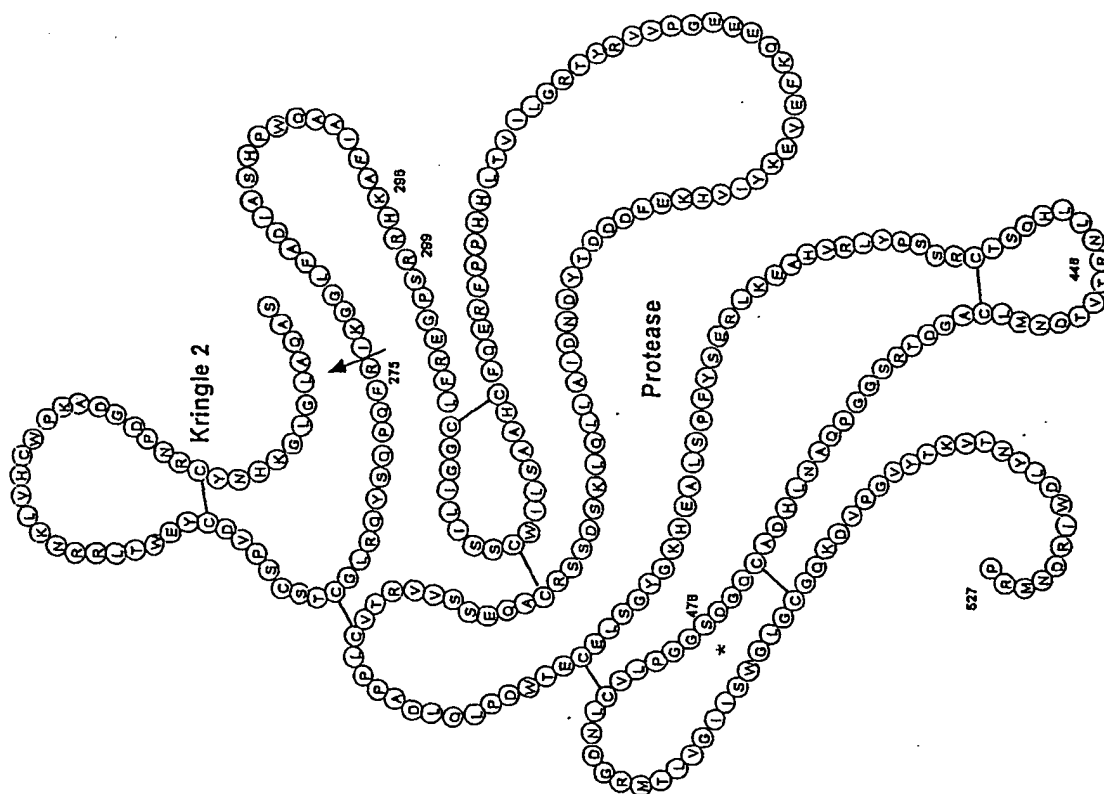


Fig. 14

14/15

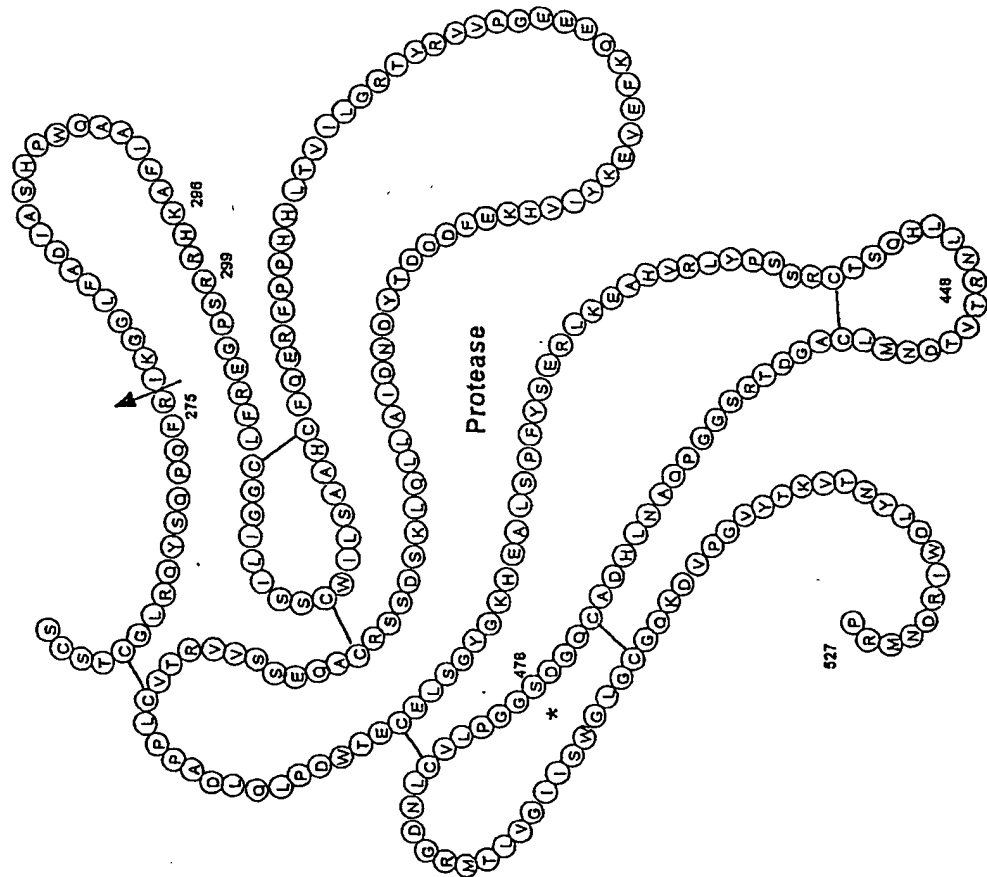


Fig. 15

15/15

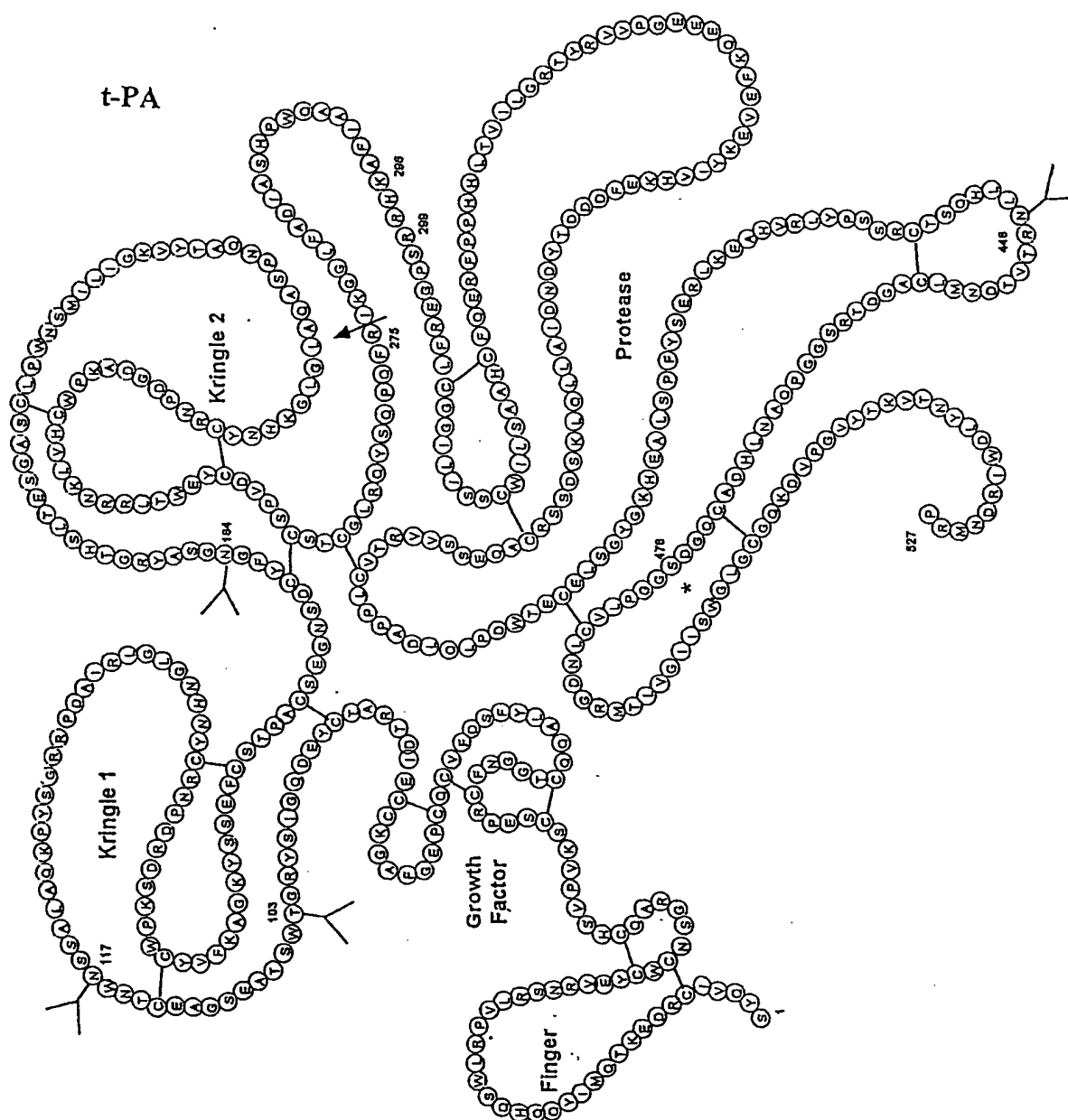


Fig. 16

Sequence Listing

<110> Boehringer Ingelheim International GmbH

<120> Methods for Large Protein Prpduction in Procaryotes

<130> sequence protocol case 1 1169

<140>

<141>

<150> GB 0027782.2

<151> 2000-11-14

<160> 18

<170> PatentIn Ver. 2.1

<210> 1 (SEQ ID NO:1)

<211> 66

<212> DNA

<213> Escherichia coli

<400> 1

atgaaaaaga cagctatcgc gattgcagtg gcactggctg gtttcgctac cgtggcccag 60
gcggcc 66

<210> 2 (SEQ ID NO:2)

<211> 4

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: N-terminal
part of K2S molecule

<400> 2

Ser Glu Gly Asn

1

<210> 3 (SEQ ID NO:3)

<211> 6

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: N-terminal
part of K2S molecule

<400> 3

Ser Glu Gly Asn Ser Asp

1

5

<210> 4 (SEQ ID NO:4)

<211> 12

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: coding

sequence of the N-terminal part of K2S molecule

<400> 4
tctgagggaa ac 12

<210> 5 (SEQ ID NO:5)
<211> 18
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: coding
sequence of the N-terminal part of K2S molecule

<400> 5
tctgagggaa acagtgc 18

<210> 6 (SEQ ID NO:6)
<211> 42
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: oligonucleotide
sequence

<400> 6
gaggaggagg tggcccaggc ggcctctgag ggaaacagtgc ac 42

<210> 7 (SEQ ID NO:7)
<211> 42
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: oligonucleotide
sequence

<400> 7
gaggaggagc tggccggcct ggcccggtcg catgttgtca cg 42

<210> 8 (SEQ ID NO:8)
<211> 26
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: oligonucleotide
sequence

<400> 8
acatgcgacc gtgacaggcc ggccag 26

<210> 9 (SEQ ID NO:9)
<211> 26
<212> DNA
<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: oligonucleotid
sequence

<400> 9

ctggccggcc tgtcacggtc gcatgt

26

<210> 10 (SEQ ID NO:10)

<211> 354

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: part of the
recombinant K2S molecule

<400> 10

Ser Glu Gly Asn Ser Asp Cys Tyr Phe Gly Asn Gly Ser Ala Tyr Arg
1 5 10 15

Gly Thr His Ser Leu Thr Glu Ser Gly Ala Ser Cys Leu Pro Trp Asn
20 25 30

Ser Met Ile Leu Ile Gly Lys Val Tyr Thr Ala Gln Asn Pro Ser Ala
35 40 45

Gln Ala Leu Gly Leu Gly Lys His Asn Tyr Cys Arg Asn Pro Asp Gly
50 55 60

Asp Ala Lys Pro Trp Cys His Val Leu Lys Asn Arg Arg Leu Thr Trp
65 70 75 80

Glu Tyr Cys Asp Val Pro Ser Cys Ser Thr Cys Gly Leu Arg Gln Tyr
85 90 95

Ser Gln Pro Gln Phe Arg Ile Lys Gly Gly Leu Phe Ala Asp Ile Ala
100 105 110

Ser His Pro Trp Gln Ala Ala Ile Phe Ala Lys His Arg Arg Ser Pro
115 120 125

Gly Glu Arg Phe Leu Cys Gly Gly Ile Leu Ile Ser Ser Cys Trp Ile
130 135 140

Leu Ser Ala Ala His Cys Phe Gln Glu Arg Phe Pro Pro His His Leu
145 150 155 160

Thr Val Ile Leu Gly Arg Thr Tyr Arg Val Val Pro Gly Glu Glu Glu
165 170 175

Gln Lys Phe Glu Val Glu Lys Tyr Ile Val His Lys Glu Phe Asp Asp
180 185 190

Asp Thr Tyr Asp Asn Asp Ile Ala Leu Leu Gln Leu Lys Ser Asp Ser
195 200 205

Ser Arg Cys Ala Gln Glu Ser Ser Val Val Arg Thr Val Cys Leu Pro
210 215 220

Pro Ala Asp Leu Gln Leu Pro Asp Trp Thr Glu Cys Glu Leu Ser Gly
225 230 235 240

Tyr Gly Lys His Glu Ala Leu Ser Pro Phe Tyr Ser Glu Arg Leu Lys
245 250 255

Glu Ala His Val Arg Leu Tyr Pro Ser Ser Arg Cys Thr Ser Gln His
 260 265 270
 5 Leu Leu Asn Arg Thr Val Thr Asp Asn Met Leu Cys Ala Gly Asp Thr
 275 280 285
 Arg Ser Gly Gly Pro Gln Ala Asn Leu His Asp Ala Cys Gln Gly Asp
 290 295 300
 10 Ser Gly Gly Pro Leu Val Cys Leu Asn Asp Gly Arg Met Thr Leu Val
 305 310 315 320
 Gly Ile Ile Ser Trp Gly Leu Gly Cys Gly Gln Lys Asp Val Pro Gly
 325 330 335
 15 Val Tyr Thr Lys Val Thr Asn Tyr Leu Asp Trp Ile Arg Asp Asn Met
 340 345 350
 20 Arg Pro

<210> 11 (SEQ ID NO:11)
 25 <211> 331
 <212> PRT
 <213> Artificial Sequence

<220>
 30 <223> Description of Artificial Sequence: part of the
 recombinant K2S molecule

<400> 11
 35 Ser Gly Ala Ser Cys Leu Pro Trp Asn Ser Met Ile Leu Ile Gly Lys
 1 5 10 15
 Val Tyr Thr Ala Gln Asn Pro Ser Ala Gln Ala Leu Gly Leu Gly Lys
 20 25 30
 40 His Asn Tyr Cys Arg Asn Pro Asp Gly Asp Ala Lys Pro Trp Cys His
 35 40 45
 Val Leu Lys Asn Arg Arg Leu Thr Trp Glu Tyr Cys Asp Val Pro Ser
 50 55 60
 45 Cys Ser Thr Cys Gly Leu Arg Gln Tyr Ser Gln Pro Gln Phe Arg Ile
 65 70 75 80
 50 Lys Gly Gly Leu Phe Ala Asp Ile Ala Ser His Pro Trp Gln Ala Ala
 85 90 95
 Ile Phe Ala Lys His Arg Arg Ser Pro Gly Glu Arg Phe Leu Cys Gly
 100 105 110
 55 Gly Ile Leu Ile Ser Ser Cys Trp Ile Leu Ser Ala Ala His Cys Phe
 115 120 125
 Gln Glu Arg Phe Pro Pro His His Leu Thr Val Ile Leu Gly Arg Thr
 130 135 140
 60 Tyr Arg Val Val Pro Gly Glu Glu Glu Gln Lys Phe Glu Val Glu Lys
 145 150 155 160
 Tyr Ile Val His Lys Glu Phe Asp Asp Asp Thr Tyr Asp Asn Asp Ile

165 170 175
 Ala Leu Leu Gln Leu Lys Ser Asp Ser Ser Arg Cys Ala Gln Glu Ser
 180 185 190
 5 Ser Val Val Arg Thr Val Cys Leu Pro Pro Ala Asp Leu Gln Leu Pro
 195 200 205
 Asp Trp Thr Glu Cys Glu Leu Ser Gly Tyr Gly Lys His Glu Ala Leu
 10 210 215 220
 Ser Pro Phe Tyr Ser Glu Arg Leu Lys Glu Ala His Val Arg Leu Tyr
 225 230 235 240
 15 Pro Ser Ser Arg Cys Thr Ser Gln His Leu Leu Asn Arg Thr Val Thr
 245 250 255
 Asp Asn Met Leu Cys Ala Gly Asp Thr Arg Ser Gly Gly Pro Gln Ala
 260 265 270
 20 Asn Leu His Asp Ala Cys Gln Gly Asp Ser Gly Gly Pro Leu Val Cys
 275 280 285
 Leu Asn Asp Gly Arg Met Thr Leu Val Gly Ile Ile Ser Trp Gly Leu
 25 290 295 300
 Gly Cys Gly Gln Lys Asp Val Pro Gly Val Tyr Thr Lys Val Thr Asn
 305 310 315 320
 30 Tyr Leu Asp Trp Ile Arg Asp Asn Met Arg Pro
 325 330

 <210> 12 (SEQ ID NO:12)
 35 <211> 339
 <212> PRT
 <213> Artificial Sequence

 <220>
 40 <223> Description of Artificial Sequence: part of the
 recombinant K2S molecule (modified)

 <400> 12
 Ser Glu Gly Asn Ser Leu Thr Glu Ser Gly Ala Ser Cys Leu Pro Trp
 45 1 5 10 15
 Asn Ser Met Ile Leu Ile Gly Lys Val Tyr Thr Ala Gln Asn Pro Ser
 20 25 30
 50 Ala Gln Ala Leu Gly Leu Gly Lys His Asn Tyr Cys Arg Asn Pro Asp
 35 40 45
 Gly Asp Ala Lys Pro Trp Cys His Val Leu Lys Asn Arg Arg Leu Thr
 50 55 60
 55 Trp Glu Tyr Cys Asp Val Pro Ser Cys Ser Thr Cys Gly Leu Arg Gln
 65 70 75 80
 Tyr Ser Gln Pro Gln Phe Arg Ile Lys Gly Gly Leu Phe Ala Asp Ile
 60 85 90 95
 Ala Ser His Pro Trp Gln Ala Ala Ile Phe Ala Lys His Arg Arg Ser
 100 105 110

Pro Gly Glu Arg Phe Leu Cys Gly Gly Ile Leu Ile Ser Ser Cys Trp
115 120 125

Ile Leu Ser Ala Ala His Cys Phe Gln Glu Arg Phe Pro Pro His His
130 135 140

Leu Thr Val Ile Leu Gly Arg Thr Tyr Arg Val Val Pro Gly Glu Glu
145 150 155 160

Glu Gln Lys Phe Glu Val Glu Lys Tyr Ile Val His Lys Glu Phe Asp
165 170 175

Asp Asp Thr Tyr Asp Asn Asp Ile Ala Leu Leu Gln Leu Lys Ser Asp
180 185 190

Ser Ser Arg Cys Ala Gln Glu Ser Ser Val Val Arg Thr Val Cys Leu
195 200 205

Pro Pro Ala Asp Leu Gln Leu Pro Asp Trp Thr Glu Cys Glu Leu Ser
210 215 220

Gly Tyr Gly Lys His Glu Ala Leu Ser Pro Phe Tyr Ser Glu Arg Leu
225 230 235 240

Lys Glu Ala His Val Arg Leu Tyr Pro Ser Ser Arg Cys Thr Ser Gln
245 250 255

His Leu Leu Asn Arg Thr Val Thr Asp Asn Met Leu Cys Ala Gly Asp
260 265 270

Thr Arg Ser Gly Gly Pro Gln Ala Asn Leu His Asp Ala Cys Gln Gly
275 280 285

Asp Ser Gly Gly Pro Leu Val Cys Leu Asn Asp Gly Arg Met Thr Leu
290 295 300

Val Gly Ile Ile Ser Trp Gly Leu Gly Cys Gly Gln Lys Asp Val Pro
305 310 315 320

Gly Val Tyr Thr Lys Val Thr Asn Tyr Leu Asp Trp Ile Arg Asp Asn
325 330 335

Met Arg Pro

<210> 13 (SEQ ID NO:13)

<211> 335

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: part of the
recombinant K2S molecule (modified)

<400> 13

Ser Leu Thr Glu Ser Gly Ala Ser Cys Leu Pro Trp Asn Ser Met Ile
1 5 10 15

Leu Ile Gly Lys Val Tyr Thr Ala Gln Asn Pro Ser Ala Gln Ala Leu
20 25 30

Gly Leu Gly Lys His Asn Tyr Cys Arg Asn Pro Asp Gly Asp Ala Lys
35 40 45

Pro Trp Cys His Val Leu Lys Asn Arg Arg Leu Thr Trp Glu Tyr Cys
 50 55 60
 5 Asp Val Pro Ser Ser Ser Thr Cys Gly Leu Arg Gln Tyr Ser Gln Pro
 65 70 75 80
 Gln Phe Arg Ile Lys Gly Gly Leu Phe Ala Asp Ile Ala Ser His Pro
 85 90 95
 10 Trp Gln Ala Ala Ile Phe Ala Lys His Arg Arg Ser Pro Gly Glu Arg
 100 105 110
 Phe Leu Cys Gly Gly Ile Leu Ile Ser Ser Cys Trp Ile Leu Ser Ala
 115 120 125
 15 Ala His Cys Phe Gln Glu Arg Phe Pro Pro His His Leu Thr Val Ile
 130 135 140
 20 Leu Gly Arg Thr Tyr Arg Val Val Pro Gly Glu Glu Glu Gln Lys Phe
 145 150 155 160
 Glu Val Glu Lys Tyr Ile Val His Lys Glu Phe Asp Asp Asp Thr Tyr
 165 170 175
 25 Asp Asn Asp Ile Ala Leu Leu Gln Leu Lys Ser Asp Ser Ser Arg Cys
 180 185 190
 Ala Gln Glu Ser Ser Val Val Arg Thr Val Cys Leu Pro Pro Ala Asp
 195 200 205
 30 Leu Gln Leu Pro Asp Trp Thr Glu Cys Glu Leu Ser Gly Tyr Gly Lys
 210 215 220
 35 His Glu Ala Leu Ser Pro Phe Tyr Ser Glu Arg Leu Lys Glu Ala His
 225 230 235 240
 Val Arg Leu Tyr Pro Ser Ser Arg Cys Thr Ser Gln His Leu Leu Asn
 245 250 255
 40 Arg Thr Val Thr Asp Asn Met Leu Cys Ala Gly Asp Thr Arg Ser Gly
 260 265 270
 Gly Pro Gln Ala Asn Leu His Asp Ala Cys Gln Gly Asp Ser Gly Gly
 275 280 285
 45 Pro Leu Val Cys Leu Asn Asp Gly Arg Met Thr Leu Val Gly Ile Ile
 290 295 300
 50 Ser Trp Gly Leu Gly Cys Gly Gln Lys Asp Val Pro Gly Val Tyr Thr
 305 310 315 320
 Lys Val Thr Asn Tyr Leu Asp Trp Ile Arg Asp Asn Met Arg Pro
 325 330 335
 55

<210> 14 (SEQ ID NO:14)

<211> 343

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: part of the recombinant K2S molecule (modified)

<400> 14

Ser Glu Gly Asn Ser Asp Thr His Ser Leu Thr Glu Ser Gly Ala Ser
 1 5 10 15
 5 Cys Leu Pro Trp Asn Ser Met Ile Leu Ile Gly Lys Val Tyr Thr Ala
 20 25 30
 Gln Asn Pro Ser Ala Gln Ala Leu Gly Leu Gly Lys His Asn Tyr Cys
 10 35 40 45
 Arg Asn Pro Asp Gly Asp Ala Lys Pro Trp Cys His Val Leu Lys Asn
 50 55 60
 15 Arg Arg Leu Thr Trp Glu Tyr Cys Asp Val Pro Ser Cys Ser Thr Cys
 65 70 75 80
 Gly Leu Arg Gln Tyr Ser Gln Pro Gln Phe Arg Ile Lys Gly Gly Leu
 85 90 95
 20 Phe Ala Asp Ile Ala Ser His Pro Trp Gln Ala Ala Ile Phe Ala Lys
 100 105 110
 His Arg Arg Ser Pro Gly Glu Arg Phe Leu Cys Gly Gly Ile Leu Ile
 115 120 125
 25 Ser Ser Cys Trp Ile Leu Ser Ala Ala His Cys Phe Gln Glu Arg Phe
 130 135 140
 30 Pro Pro His His Leu Thr Val Ile Leu Gly Arg Thr Tyr Arg Val Val
 145 150 155 160
 Pro Gly Glu Glu Glu Gln Lys Phe Glu Val Glu Lys Tyr Ile Val His
 165 170 175
 35 Lys Glu Phe Asp Asp Asp Thr Tyr Asp Asn Asp Ile Ala Leu Leu Gln
 180 185 190
 Leu Lys Ser Asp Ser Ser Arg Cys Ala Gln Glu Ser Ser Val Val Arg
 195 200 205
 40 Thr Val Cys Leu Pro Pro Ala Asp Leu Gln Leu Pro Asp Trp Thr Glu
 210 215 220
 45 Cys Glu Leu Ser Gly Tyr Gly Lys His Glu Ala Leu Ser Pro Phe Tyr
 225 230 235 240
 Ser Glu Arg Leu Lys Glu Ala His Val Arg Leu Tyr Pro Ser Ser Arg
 245 250 255
 50 Cys Thr Ser Gln His Leu Leu Asn Arg Thr Val Thr Asp Asn Met Leu
 260 265 270
 Cys Ala Gly Asp Thr Arg Ser Gly Gly Pro Gln Ala Asn Leu His Asp
 275 280 285
 55 Ala Cys Gln Gly Asp Ser Gly Gly Pro Leu Val Cys Leu Asn Asp Gly
 290 295 300
 60 Arg Met Thr Leu Val Gly Ile Ile Ser Trp Gly Leu Gly Cys Gly Gln
 305 310 315 320
 Lys Asp Val Pro Gly Val Tyr Thr Lys Val Thr Asn Tyr Leu Asp (Trp
 325 330 335

Ile Arg Asp Asn Met Arg Pro
340

<210> 15 (SEQ ID NO:15)
<211> 343
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: part of the
recombinant K2S molecule (modified)

<400> 15
Ser Glu Gly Asn Ser Asp Thr His Ser Leu Thr Glu Ser Gly Ala Ser
1 5 10 15
Cys Leu Pro Trp Asn Ser Met Ile Leu Ile Gly Lys Val Tyr Thr Ala
20 25 30
Gln Asn Pro Ser Ala Gln Ala Leu Gly Leu Gly Lys His Asn Tyr Cys
35 40 45
Arg Asn Pro Asp Gly Asp Ala Lys Pro Trp Cys His Val Leu Lys Asn
50 55 60
Arg Arg Leu Thr Trp Glu Tyr Cys Asp Val Pro Ser Ser Ser Thr Cys
65 70 75 80
Gly Leu Arg Gln Tyr Ser Gln Pro Gln Phe Arg Ile Lys Gly Gly Leu
85 90 95
Phe Ala Asp Ile Ala Ser His Pro Trp Gln Ala Ala Ile Phe Ala Lys
100 105 110
His Arg Arg Ser Pro Gly Glu Arg Phe Leu Cys Gly Gly Ile Leu Ile
115 120 125
Ser Ser Cys Trp Ile Leu Ser Ala Ala His Cys Phe Gln Glu Arg Phe
130 135 140
Pro Pro His His Leu Thr Val Ile Leu Gly Arg Thr Tyr Arg Val Val
145 150 155 160
Pro Gly Glu Glu Glu Gln Lys Phe Glu Val Glu Lys Tyr Ile Val His
165 170 175
Lys Glu Phe Asp Asp Asp Thr Tyr Asp Asn Asp Ile Ala Leu Leu Gln
180 185 190
Leu Lys Ser Asp Ser Ser Arg Cys Ala Gln Glu Ser Ser Val Val Arg
195 200 205
Thr Val Cys Leu Pro Pro Ala Asp Leu Gln Leu Pro Asp Trp Thr Glu
210 215 220
Cys Glu Leu Ser Gly Tyr Gly Lys His Glu Ala Leu Ser Pro Phe Tyr
225 230 235 240
Ser Glu Arg Leu Lys Glu Ala His Val Arg Leu Tyr Pro Ser Ser Arg
245 250 255
Cys Thr Ser Gln His Leu Leu Asn Arg Thr Val Thr Asp Asn Met Leu

```

      260      265      270
Cys Ala Gly Asp Thr Arg Ser Gly Gly Pro Gln Ala Asn Leu His Asp
      275      280      285
5  Ala Cys Gln Gly Asp Ser Gly Gly Pro Leu Val Cys Leu Asn Asp Gly
      290      295      300
10 Arg Met Thr Leu Val Gly Ile Ile Ser Trp Gly Leu Gly Cys Gly Gln
      305      310      315      320
15 Lys Asp Val Pro Gly Val Tyr Thr Lys Val Thr Asn Tyr Leu Asp Trp
      325      330      335
      Ile Arg Asp Asn Met Arg Pro
      340

<210> 16 (SEQ ID NO:16)
20 <211> 308
    <212> PRT
    <213> Artificial Sequence

    <220>
25 <223> Description of Artificial Sequence: part of the
    recombinant K2S molecule (modified)

    <400> 16
30 Ser Ala Gln Ala Leu Gly Leu Gly Lys His Asn Tyr Cys Arg Asn Pro
      1      5      10      15
    Asp Gly Asp Ala Lys Pro Trp Cys His Val Leu Lys Asn Arg Arg Leu
      20      25      30
35 Thr Trp Glu Tyr Cys Asp Val Pro Ser Cys Ser Thr Cys Gly Leu Arg
      35      40      45
    Gln Tyr Ser Gln Pro Gln Phe Arg Ile Lys Gly Gly Leu Phe Ala Asp
      50      55      60
40 Ile Ala Ser His Pro Trp Gln Ala Ala Ile Phe Ala Lys His Arg Arg
      65      70      75      80
    Ser Pro Gly Glu Arg Phe Leu Cys Gly Gly Ile Leu Ile Ser Ser Cys
45      85      90      95
    Trp Ile Leu Ser Ala Ala His Cys Phe Gln Glu Arg Phe Pro Pro His
      100      105      110
50 His Leu Thr Val Ile Leu Gly Arg Thr Tyr Arg Val Val Pro Gly Glu
      115      120      125
    Glu Glu Gln Lys Phe Glu Val Glu Lys Tyr Ile Val His Lys Glu Phe
      130      135      140
55 Asp Asp Asp Thr Tyr Asp Asn Asp Ile Ala Leu Leu Gln Leu Lys Ser
      145      150      155      160
    Asp Ser Ser Arg Cys Ala Gln Glu Ser Ser Val Val Arg Thr Val Cys
60      165      170      175
    Leu Pro Pro Ala Asp Leu Gln Leu Pro Asp Trp Thr Glu Cys Glu Leu
      180      185      190

```

Ser Gly Tyr Gly Lys His Glu Ala Leu Ser Pro Phe Tyr Ser Glu Arg
 195 200 205
 Leu Lys Glu Ala His Val Arg Leu Tyr Pro Ser Ser Arg Cys Thr Ser
 210 215 220
 Gln His Leu Leu Asn Arg Thr Val Thr Asp Asn Met Leu Cys Ala Gly
 225 230 235 240
 Asp Thr Arg Ser Gly Gly Pro Gln Ala Asn Leu His Asp Ala Cys Gln
 245 250 255
 Gly Asp Ser Gly Gly Pro Leu Val Cys Leu Asn Asp Gly Arg Met Thr
 260 265 270
 Leu Val Gly Ile Ile Ser Trp Gly Leu Gly Cys Gly Gln Lys Asp Val
 275 280 285
 Pro Gly Val Tyr Thr Lys Val Thr Asn Tyr Leu Asp Trp Ile Arg Asp
 290 295 300
 Asn Met Arg Pro
 305
 <210> 17 (SEQ ID NO:17)
 <211> 268
 <212> PRT
 <213> Artificial Sequence
 <220>
 <223> Description of Artificial Sequence: part of the
 recombinant K2S molecule (modified)
 <400> 17
 Ser Cys Ser Thr Cys Gly Leu Arg Gln Tyr Ser Gln Pro Gln Phe Arg
 1 5 10 15
 Ile Lys Gly Gly Leu Phe Ala Asp Ile Ala Ser His Pro Trp Gln Ala
 20 25 30
 Ala Ile Phe Ala Lys His Arg Arg Ser Pro Gly Glu Arg Phe Leu Cys
 35 40 45
 Gly Gly Ile Leu Ile Ser Ser Cys Trp Ile Leu Ser Ala Ala His Cys
 50 55 60
 Phe Gln Glu Arg Phe Pro Pro His His Leu Thr Val Ile Leu Gly Arg
 65 70 75 80
 Thr Tyr Arg Val Val Pro Gly Glu Glu Gln Lys Phe Glu Val Glu
 85 90 95
 Lys Tyr Ile Val His Lys Glu Phe Asp Asp Asp Thr Tyr Asp Asn Asp
 100 105 110
 Ile Ala Leu Leu Gln Leu Lys Ser Asp Ser Ser Arg Cys Ala Gln Glu
 115 120 125
 Ser Ser Val Val Arg Thr Val Cys Leu Pro Pro Ala Asp Leu Gln Leu
 130 135 140
 Pro Asp Trp Thr Glu Cys Glu Leu Ser Gly Tyr Gly Lys His Glu Ala
 145 150 155 160

12/13

Leu Ser Pro Phe Tyr Ser Glu Arg Leu Lys Glu Ala His Val Arg Leu
 165 170 175
 Tyr Pro Ser Ser Arg Cys Thr Ser Gln His Leu Leu Asn Arg Thr Val
 180 185 190
 Thr Asp Asn Met Leu Cys Ala Gly Asp Thr Arg Ser Gly Gly Pro Gln
 195 200 205
 Ala Asn Leu His Asp Ala Cys Gln Gly Asp Ser Gly Gly Pro Leu Val
 210 215 220
 Cys Leu Asn Asp Gly Arg Met Thr Leu Val Gly Ile Ile Ser Trp Gly
 225 230 235 240
 Leu Gly Cys Gly Gln Lys Asp Val Pro Gly Val Tyr Thr Lys Val Thr
 245 250 255
 Asn Tyr Leu Asp Trp Ile Arg Asp Asn Met Arg Pro
 260 265
 <210> 18 (SEQ ID NO:18)
 <211> 527
 <212> PRT
 <213> Homo sapiens (tPA)
 <400> 18
 Ser Tyr Gln Val Ile Cys Arg Asp Glu Lys Thr Gln Met Ile Tyr Gln
 1 5 10 15
 Gln His Gln Ser Trp Leu Arg Pro Val Leu Arg Ser Asn Arg Val Glu
 20 25 30
 Tyr Cys Trp Cys Asn Ser Gly Arg Ala Gln Cys His Ser Val Pro Val
 35 40 45
 Lys Ser Cys Ser Glu Pro Arg Cys Phe Asn Gly Gly Thr Cys Gln Gln
 50 55 60
 Ala Leu Tyr Phe Ser Asp Phe Val Cys Gln Cys Pro Glu Gly Phe Ala
 65 70 75 80
 Gly Lys Cys Cys Glu Ile Asp Thr Arg Ala Thr Cys Tyr Glu Asp Gln
 85 90 95
 Gly Ile Ser Tyr Arg Gly Thr Trp Ser Thr Ala Glu Ser Gly Ala Glu
 100 105 110
 Cys Thr Asn Trp Asn Ser Ser Ala Leu Ala Gln Lys Pro Tyr Ser Gly
 115 120 125
 Arg Arg Pro Asp Ala Ile Arg Leu Gly Leu Gly Asn His Asn Tyr Cys
 130 135 140
 Arg Asn Pro Asp Arg Asp Ser Lys Pro Trp Cys Tyr Val Phe Lys Ala
 145 150 155 160
 Gly Lys Tyr Ser Ser Glu Phe Cys Ser Thr Pro Ala Cys Ser Glu Gly
 165 170 175
 Asn Ser Asp Cys Tyr Phe Gly Asn Gly Ser Ala Tyr Arg Gly Thr His
 180 185 190

Ser Leu Thr Glu Ser Gly Ala Ser Cys Leu Pro Trp Asn Ser Met Ile
 195 200 205
 5 Leu Ile Gly Lys Val Tyr Thr Ala Gln Asn Pro Ser Ala Gln Ala Leu
 210 215 220
 Gly Leu Gly Lys His Asn Tyr Cys Arg Asn Pro Asp Gly Asp Ala Lys
 225 230 235 240
 10 Pro Trp Cys His Val Leu Lys Asn Arg Arg Leu Thr Trp Glu Tyr Cys
 245 250 255
 Asp Val Pro Ser Cys Ser Thr Cys Gly Leu Arg Gln Tyr Ser Gln Pro
 15 260 265 270
 Gln Phe Arg Ile Lys Gly Gly Leu Phe Ala Asp Ile Ala Ser His Pro
 275 280 285
 20 Trp Gln Ala Ala Ile Phe Ala Lys His Arg Arg Ser Pro Gly Glu Arg
 290 295 300
 Phe Leu Cys Gly Gly Ile Leu Ile Ser Ser Cys Trp Ile Leu Ser Ala
 305 310 315 320
 25 Ala His Cys Phe Gln Glu Arg Phe Pro Pro His His Leu Thr Val Ile
 325 330 335
 Leu Gly Arg Thr Tyr Arg Val Val Pro Gly Glu Glu Glu Gln Lys Phe
 30 340 345 350
 Glu Val Glu Lys Tyr Ile Val His Lys Glu Phe Asp Asp Asp Thr Tyr
 355 360 365
 35 Asp Asn Asp Ile Ala Leu Leu Gln Leu Lys Ser Asp Ser Ser Arg Cys
 370 375 380
 Ala Gln Glu Ser Ser Val Val Arg Thr Val Cys Leu Pro Pro Ala Asp
 385 390 395 400
 40 Leu Gln Leu Pro Asp Trp Thr Glu Cys Glu Leu Ser Gly Tyr Gly Lys
 405 410 415
 His Glu Ala Leu Ser Pro Phe Tyr Ser Glu Arg Leu Lys Glu Ala His
 45 420 425 430
 Val Arg Leu Tyr Pro Ser Ser Arg Cys Thr Ser Gln His Leu Leu Asn
 435 440 445
 50 Arg Thr Val Thr Asp Asn Met Leu Cys Ala Gly Asp Thr Arg Ser Gly
 450 455 460
 Gly Pro Gln Ala Asn Leu His Asp Ala Cys Gln Gly Asp Ser Gly Gly
 465 470 475 480
 55 Pro Leu Val Cys Leu Asn Asp Gly Arg Met Thr Leu Val Gly Ile Ile
 485 490 495
 Ser Trp Gly Leu Gly Cys Gly Gln Lys Asp Val Pro Gly Val Tyr Thr
 60 500 505 510
 Lys Val Thr Asn Tyr Leu Asp Trp Ile Arg Asp Asn Met Arg Pro
 515 520 525